

Crops and Soils Research Paper

Cite this article: Obeng-Bio E, Badu-Apraku B, Ifie BE, Danquah A, Blay ET, Annor B (2019). Genetic analysis of grain yield and agronomic traits of early provitamin A quality protein maize inbred lines in contrasting environments. *The Journal of Agricultural Science* **157**, 413–433. <https://doi.org/10.1017/S0021859619000753>

Received: 27 April 2019

Revised: 9 September 2019

Accepted: 24 September 2019

Key words:

Drought tolerance; low-N tolerance; provitamin A; quality protein maize; *Zea mays* L.

Author for correspondence:

B. Badu-Apraku, E-mail: b.badu-apraku@cgiar.org

Genetic analysis of grain yield and agronomic traits of early provitamin A quality protein maize inbred lines in contrasting environments

E. Obeng-Bio^{1,2}, B. Badu-Apraku³ , B. E. Ifie¹, A. Danquah¹, E. T. Blay¹ and B. Annor²

¹West Africa Center for Crop Improvement (WACCI), University of Ghana, PBM 30, Legon, Accra, Ghana; ²CSIR-Crops Research Institute, P. O. Box 3785, Fumesua, Kumasi, Ghana and ³International Institute of Tropical Agriculture (IITA), PMB 5320, Oyo Rd, Ibadan, Nigeria

Abstract

Early-maturing provitamin A (PVA) quality protein maize (QPM) hybrids with combined drought and low soil nitrogen (low-N) tolerance are needed to address malnutrition and food security problems in sub-Saharan Africa (SSA). The current study's objectives were to (i) examine combining ability of selected early maturing PVA-QPM inbreds for grain yield and other agronomic traits under drought, low-N, optimal environments and across environments, (ii) determine gene action conditioning PVA accumulation under optimal environments, (iii) classify inbreds into heterotic groups and identify testers and (iv) assess yield and stability of hybrids across environments. Ninety-six hybrids generated from 24 inbred lines using the North Carolina Design II together with four commercial hybrid controls were evaluated under drought, low-N and optimal environments in Nigeria in 2016 and 2017. Fifty-four selected hybrids were assayed for PVA carotenoid and tryptophan content. Additive genetic effects were greater than non-additive effects for grain yield and most agronomic traits under each and across environments. The gene action conditioning accumulation of PVA carotenoids under optimal growing conditions followed a pattern similar to that of grain yield and other yield-related traits. The inbred lines were categorized into four heterotic groups consistent with the pedigree records and with TZEIORQ 29 identified as the best male and female tester for heterotic group IV. No tester was found for the other groups. Hybrid TZEIORQ 24 × TZEIORQ 41 was the highest yielding and most stable across environments and should be further tested for consistent performance for commercialization in SSA.

Introduction

Maize has the greatest potential for increased production and productivity in the savannah belt of sub-Saharan Africa (SSA) due to high solar radiation, low night temperatures and low incidence of pests and diseases that characterize the region. Despite this potential, grain yield of maize in farmers' fields has been consistently low, with an average of 1.7 t/ha (FAO, 2016). This challenge has been ascribed to several factors including *Striga hermonthica* infestation, drought and low nitrogen (N).

The response of maize to drought and low-N has been found to be influenced by similar mechanisms (Badu-Apraku *et al.*, 2012) and the two stresses together cause significant yield reduction in SSA (Badu-Apraku *et al.*, 2011a). Drought at flowering and grain filling periods may cause losses of 40–90% (Menkir and Akintunde, 2001). Grain yield losses resulting from low-N, on the other hand, could vary between 10 and 50% (Wolfe *et al.*, 1988). Improving maize to tolerate drought and low-N conditions is crucial to the ongoing efforts to reduce the yield gap between optimal and stress conditions. Edmeades (2013) found that about 0.20–0.25 of the yield gap between potential yield and moisture-limited yield could be eliminated by genetic improvement for drought and low-N tolerance.

Vitamin A is essentially needed by human beings for improved eyesight and enhanced immune system. Unfortunately, the requirement for vitamin A in humans is met through external sources because the human body is unable to synthesize this important micronutrient. Provitamin A (PVA) maize has the potential to supply more than the daily dietary requirement of 15.0 µg/g dry weight (DW) PVA for human beings as compared to about 2.0 µg/g DW available in the commonly cultivated and consumed yellow maize cultivars (Pixley *et al.*, 2013). Efforts at improving the PVA content in maize have resulted in the discovery of three genes [phytoene synthase1 (*PSY1*), lycopene epsilon cyclase (*LcyE*) and β-carotene hydroxylase1 (*crTRB1*)] that regulate the key steps involved in the synthesis and accruing of PVA carotenoids (Wurtzel *et al.*, 2012). The allele *crTRB1* (–5'TE and 3'TE) has been the most functional for increased β-carotene contents (Yan *et al.*, 2010).

© The Author(s) 2019. This is an Open Access article, distributed under the terms of the Creative Commons Attribution licence (<http://creativecommons.org/licenses/by/4.0/>), which permits unrestricted re-use, distribution, and reproduction in any medium, provided the original work is properly cited.

Conventional maize has inadequate levels of tryptophan and lysine and therefore its consumption without a balanced protein source, especially by infants, could result in initial growth failures such as 'kwashiorkor', reduced immune system and consequently, death (Sultana *et al.*, 2019). On the contrary, quality protein maize (QPM) has the potential to supply about 0.73 of human protein requirements compared to about 0.46 from conventional maize (Badu-Apraku and Fakorede, 2017). Studies aimed at elevating the protein content of maize have resulted in identification of the recessive homozygous allele of the opaque-2 gene (*o2o2*) and its modifiers, which are capable of doubling the tryptophan and lysine contents of QPM relative to their conventional counterparts (Ma and Nelson, 1975; Crow and Kermicle, 2002; Krivanek *et al.*, 2007; Badu-Apraku *et al.*, 2016).

Through the concerted efforts of the International Institute of Tropical Agriculture Maize Improvement Programme (IITA-MIP), PVA-QPM inbred lines with drought and low-N tolerance genetic backgrounds have been developed over the years to produce hybrids and synthetic varieties. Although several studies on the combining ability using different mating designs and heterotic grouping of maize inbred lines under contrasting environments have been conducted in SSA (Menkir *et al.*, 2003; Badu-Apraku *et al.*, 2013, 2015; Annor and Badu-Apraku, 2016), it is still very important to conduct such studies for newly developed inbred lines because there have been inconsistent reports on the type of gene action conditioning the inheritance of grain yield and other agronomic traits in maize under drought and low-N conditions. Furthermore, unlike lysine and tryptophan which have been widely reported to be influenced by maternal and additive genetic effects (Ngaboyisonga *et al.*, 2009; Badu-Apraku and Fakorede, 2017; Varadaraju and Joel, 2017), there have been inconsistent reports on the gene action controlling the inheritance of PVA accumulation. Several workers have reported the preponderance of additive over non-additive genetic effects in determining PVA concentrations in maize (Egesel *et al.*, 2003; Suwarno, 2012; Menkir *et al.*, 2014; Owens *et al.*, 2014; Suwarno *et al.*, 2015), suggesting that using recurrent selection methods for the improvement of PVA content would be successful (Coors, 1999). Furthermore, Menkir *et al.* (2008) and Owens *et al.* (2014) reported positive correlations among individual carotenoids while Menkir *et al.* (2014) as well as Suwarno *et al.* (2015) found positive relationships between PVA content and grain yield. These findings suggested that simultaneous increases in PVA carotenoids and grain yield may be effective (Bouis and Welch, 2010; Menkir *et al.*, 2014). On the contrary, Halilu *et al.* (2016) reported non-additive gene action to be more important in controlling the PVA carotenoid concentrations. They also found no significant genotypic correlations between carotenoid content and grain yield, suggesting that the two traits could be improved independently (Halilu *et al.*, 2016).

Assessing the combining ability of the panel of newly developed IITA early (90–95 days to physiological maturity) inbred lines is necessary to guide breeding strategies under low-N, drought and optimal growing environments. Classifying inbred lines into different heterotic groups will help to reduce the development and evaluation of less productive crosses, while exploiting maximum heterosis by crossing opposing inbred lines (Terron *et al.*, 1997). The current study was designed to (i) examine the combining ability of selected early maturing PVA-QPM inbred lines for grain yield and other traits under low-N, drought and optimal conditions, and across environments, (ii) determine the gene action conditioning PVA accumulation under optimal growing environments, (iii)

categorize the inbred lines into heterotic groups and identify the best testers across the contrasting environments and (iv) assess the grain yield and stability of the single crosses across test environments.

Materials and methods

Genetic materials and development of inbred lines and hybrids

The 24 early-maturing PVA-QPM inbred lines used in the study were developed by the IITA-MIP (Table 1). The inbred lines were selected based on their reactions to drought and low-N environments in previous studies (Badu-Apraku and Fakorede, 2017). For the PVA and quality protein traits, kernel colour and endosperm modification, respectively, were used to select parental lines. The 24 parental inbreds were grouped into six sets with each set containing four inbred lines that were used to generate 96 single cross hybrids, employing the North Carolina Design II (NCD II) method proposed by Comstock and Robinson (1948) during the 2015/2016 dry season at IITA, Ibadan, Nigeria.

Field evaluations under the different environmental conditions

The 96 PVA-QPM single cross hybrids plus four controls were assessed under drought, low-N and optimal growing conditions in Nigeria, for 2 years each. The managed drought experiments were conducted in the dry seasons at Ikenne (forest-savannah transitional zone, 7°52'N, 30°44'E, 61 m a.s.l., 1200 mm mean annual precipitation) from November 2016 to February 2017 and at Kadawa (semi-arid Sudan savannah, 12°45'N, 9°45'E, 469 m a.s.l., 884 mm mean annual precipitation) from February to May 2018. The managed drought condition at Ikenne was attained by providing 17 mm of irrigation water every week from sowing until 28 days after planting (DAP), when the irrigation was terminated and the hybrids depended on stored soil moisture to reach physiological maturity. Nitrogen-phosphorus-potassium (NPK at 15 : 15 : 15) fertilizer was applied to the managed drought trials at the rate of 60 kg/ha each of N, P and K at planting. Additionally, 60 kg/ha of urea was applied 2 weeks after planting. The managed drought at Kadawa was achieved using a furrow irrigation system that also provided water once per week up to 28 DAP, when the irrigation was terminated. However, irrigation was resumed 2 weeks after flowering to avoid total loss of the trial due to the intense heat that characterizes Kadawa between February and March and the characteristic sandy and shallow top soil of the test environment.

The hybrids were also evaluated under low-N (30 kg/ha) environments at Ile-Ife (forest-savannah transition, 8°28'N, 5°34'E, and 245 m a.s.l., 1300 mm mean annual precipitation) and Mokwa (Southern Guinea savannah, 9°20'N, 5°5'E, 458 m a.s.l., 1100 mm mean annual precipitation) during the 2016 and 2017 growing seasons. The experimental fields at both locations were depleted of N by continuously sowing densely populated maize and removing all crop residues after harvest for three seasons. Before the low-N fields were prepared, soil samples from 0 to 15 cm depth were taken for analysis using the colorimetric and Kjeldahl digestion method (Bremner and Mulvaney, 1982) to determine the levels of N, P and K. The soil from the low-N experimental field at Mokwa contained 0.85 g/kg of N, 0.006 g/kg of P and 0.20 g/kg of K, while that of Ile-Ife had 0.84 g/kg of N, 0.002 g/kg of P and 0.358 g/kg of K. Based on the soil tests, NPK fertilizer was formulated using urea, single-superphosphate and

Table 1. Description of the 24 early maturing PVA-QPM inbreds selected for the NCD II crosses in the 2015/16 dry season at IITA Ibadan, Nigeria

Sl. no.	Inbred	Pedigree	Reaction to drought	Reaction to low-N	Set
1	TZEIORQ 69	2009-TZE OR2 DT STR QPM S ₆ inb 57-2/2-2/2-1/1-1/2-1/1	Tolerant	Tolerant	A
2	TZEIORQ 29	2009-TZE OR2 DT STR QPM S ₆ inb 28-1/1-2/2-1/2-1/2-1/1	Tolerant	Susceptible	A
3	TZEIORQ 45	2009-TZE OR2 DT STR QPM S ₆ inb 35-2/3-3/3-4/4-3/4-1/1	Susceptible	Tolerant	A
4	TZEIORQ 48	2009-TZE OR2 DT STR QPM S ₆ inb 41-1/2-1/3-1/2-3/3-1/1	Tolerant	Susceptible	A
5	TZEIORQ 11	2009-TZE OR2 DT STR QPM S ₆ inb 7-1/3-1/2-1/2-4/4-1/1	Tolerant	Tolerant	B
6	TZEIORQ 20	2009-TZE OR2 DT STR QPM S ₆ inb 26-1/1-1/2-1/6-1/2-1/1	Tolerant	Tolerant	B
7	TZEIORQ 6	2009-TZE OR2 DT STR QPM S ₆ inb 2-2/3-2/3-2/4-1/5-1/1	Tolerant	Susceptible	B
8	TZEIORQ 44	2009-TZE OR2 DT STR-QPM S ₆ inb 35-2/3-3/3-4/4-1/4-1/1	Tolerant	Susceptible	B
9	TZEIORQ 42	2009-TZE OR2 DT STR QPM S ₆ inb 35-2/3-3/3-2/4-2/2-1/1	Tolerant	Tolerant	C
10	TZEIORQ 59	2009-TZE OR2 DT STR QPM S ₆ inb 50-2/2-1/3-2/3-2/2-1/1	Tolerant	Tolerant	C
11	TZEIORQ 15	2009-TZE OR2 DT STR QPM S ₆ inb 7-2/3-1/2-3/4-1/3-1/1	Tolerant	Susceptible	C
12	TZEIORQ 23	2009-TZE OR2 DT STR QPM S ₆ inb 26-1/1-1/2-4/6-1/3-1/1	Tolerant	Susceptible	C
13	TZEIQI 82	TZE-COMP5-Y C6S6 Inb 25 × Pool 18 SR QPM BC1S6 2-3-1-1-6-6	Tolerant	Tolerant	D
14	TZEIORQ 47	2009-TZE OR2 DT STR-QPM S ₆ inb 35-3/3-3/3-1/3-2/2-1/1	Susceptible	Tolerant	D
15	TZEIORQ 7	2009-TZE OR2 DT STR-QPM S ₆ inb 2-2/3-2/3-3/4-1/3-1/1	Tolerant	Susceptible	D
16	TZEIORQ 13	2009-TZE OR2 DT STR-QPM S ₆ inb 7-1/3-1/2-2/2-3/3-1/1	Tolerant	Susceptible	D
17	TZEIORQ 2	2009-TZE OR2 DT STR-QPM S ₆ inb 2-2/3-1/3-1/3-1/2-1/1-	Tolerant	Tolerant	E
18	TZEIORQ 5	2009-TZE OR2 DT STR-QPM S ₆ inb 2-2/3-2/3-1/4-3/3-1/1	Tolerant	Susceptible	E
19	TZEIORQ 26	2009-TZE OR2 DT STR-QPM S ₆ inb 26-1/1-1/2-6/6-2/3-1/1	Tolerant	Tolerant	E
20	TZEIORQ 41	2009-TZE OR2 DT STR-QPM S ₆ inb 35-2/3-3/3-4/4-1/4-1/1	Tolerant	Susceptible	E
21	TZEIORQ 24	2009-TZE OR2 DT STR-QPM S ₆ inb 26-1/1-1/2-4/6-2/3-1/1	Tolerant	Tolerant	F
22	TZEIORQ 43	2009-TZE OR2 DT STR-QPM S ₆ inb 35-2/3-3/3-3/4-1/2-1/1	Tolerant	Susceptible	F
23	TZEIORQ 40	2009-TZE OR2 DT STR-QPM S ₆ inb 35-2/3-2/3-1/2-2/2-1/1	Susceptible	Tolerant	F
24	TZEIORQ 70	2009-TZE OR2 DT STR-QPM S ₆ inb 60-2/2-1/2-1/3-1/4-1/1	Tolerant	Susceptible	F

muriate of potash, respectively, which was applied at 14 DAP immediately after thinning to bring the levels of the total available basal N to 15 kg/ha. The levels of the single superphosphate and muriate of potash fertilizers applied were 60 kg/ha each of P and K. Additional 15 kg/ha of urea was applied at 28 DAP to bring the total available N to 30 kg/ha. Hybrid evaluations were carried out under optimal growing conditions at Ile-Ife, Mokwa and Ikenne during the 2016 and 2017 growing seasons. The soil at Ikenne and Ile-Ife is Alfisol, while that of Kadawa and Mokwa is Eutric Gambisol and Luvisol, respectively (Soil Survey Staff, 1999).

For the experiments planted under optimal conditions, the NPK (15 : 15 : 15) fertilizer was applied at 14 DAP to make available 60 kg/ha each of N, P and K. Additionally 30 kg N/ha was used for the top dressing at 28 DAP. A 10 × 10 alpha lattice design, replicated twice, was used for all hybrid experiments. Single-row plots each 4 m long, with a spacing of 0.75 m between rows and 0.40 m within rows, constituted an experimental unit. A plant population density of about 66 666 per ha was obtained by planting three seeds per hill and later thinning to two plants per stand. The inbred lines were also evaluated in adjacent blocks along with the hybrids in the different test environments. Each experimental unit of the inbred trials consisted of one-row plot each with a spacing of 0.75 m between rows and 0.40 m within rows. Weed control in the inbred and hybrid experiments was

achieved by the application of pre- and post-emergence herbicides (atrazine and gramoxone) at 5 litres/ha followed by manual weeding.

Data collection

Data were collected for 50% days to anthesis (DA) and silking (DS), and plant and ear heights (PLHT and EHT). Plant aspect (PASP) was rated on a scale of 1–9, where 1 represented excellent overall phenotypic appeal and 9 extremely poor overall phenotypic appeal. Ear aspect (EASP) was also scored on a scale of 1–9, where 1 denoted excellent ears with well-filled grains without any damage resulting from insect and disease attack while 9 represented plots with only one or no ears. The difference between DA and DS was calculated as anthesis–silking interval, while the number of ears per plant was obtained as the ratio of the number of ears harvested in a plot to the number of plants. The low-N and drought trials were rated for stay-green characteristic (STGR) at 70 DAP [growth stage (GS)85 = soft dough development (Zadoks *et al.*, 1974)] using a scale of 1–9 where 1 = 0–0.10 dead leaf area, and 9 = 0.81–1.00 dead leaf area. For the drought and low-N trials, grain weight per plot was measured. Grain moisture was determined and grain yield in kg/ha was subsequently calculated using the grain weight adjusted to 15% moisture

content. For the optimal trials however, a shelling percentage of 80 was assumed per plot for the hybrids and grain yield (adjusted to 15% moisture content) based on ear weight was calculated and converted to kg/ha.

Generation of kernel samples of hybrids for carotenoids and tryptophan analyses

Fifty-four selected PVA-QPM hybrids (out of the 96 hybrids generated) plus a standard QPM variety control, 'Obatanpa', were planted under well-watered growing conditions in January 2018 at IITA Ibadan, to produce kernel samples for carotenoid and tryptophan analyses. These were the hybrids that yielded as much or more than the estimated mean grain yield across environments, and 54 were used on the basis of retaining three out of four inbred lines per set in order to maintain the NCD II arrangement for genetic analysis. This was due to cost implications as quantification of the carotenoids and tryptophan was very expensive. The hybrids were planted using 1 m single row plots with spacings of 0.75 and 0.20 m. Two seeds were sown per hill and were thinned to one plant to provide a minimum of five plants per hybrid. Plants in each plot were self-pollinated to produce seed samples for chemical analysis. Ears of the self-pollinated F₁ hybrids were harvested and air-dried. Individual ears of each plot were shelled separately and an equal number of kernels from each ear were taken and bulked to represent a hybrid. Samples of 100 kernels for each hybrid were collected and analysed for carotenoid and tryptophan levels in the IITA nutritional laboratory.

Analysis of hybrids for carotenoids and tryptophan contents

The high-performance liquid chromatography method was used for the extraction and quantification of carotenoids based on the protocol described by Howe and Tanumihardjo (2006). Beta-carotene (*cis* and *trans* isomers), α -carotene, β -cryptoxanthin, zeaxanthin and lutein were quantified using external standards. Total carotenoids were computed as the sum of concentrations of α -carotene, β -carotene, lutein, zeaxanthin and β -cryptoxanthin. PVA was computed as the sum of β -carotene, and half of each of β -cryptoxanthin and α -carotene contents, since β -cryptoxanthin and α -carotene contribute half of the value of β -carotene as PVA (US Institute of Medicine, 2001). Values of all carotenoids for each sample were obtained from two independent measurements.

Furthermore, the selected hybrids were analysed for tryptophan levels in whole grain flour using the colorimetric method proposed by Hernández and Bates (1969). Tryptophan content was determined with the aid of a standard curve of a known control and values for each sample were obtained from two independent measurements. Percent tryptophan was expressed in terms of a given unit of samples in whole grain (Teklewold *et al.*, 2015).

Statistical analysis

Analysis of variance

The combination of location and year represented an environment while the low-N, drought and optimal conditions were regarded as research conditions. The NCD II analysis of variance (ANOVA) for each and across research conditions were performed on plot means for all data collected using the general linear model procedure (PROC GLM) implemented in Statistical

Analyses System, version 9.4 (SAS Institute, 2012). In the model, environments, replicates within environments and incomplete blocks within replicates \times environment interaction were considered as random factors whereas the set of hybrids was regarded as a fixed factor. Block effects on hybrid means were adjusted using the lattice design proposed by Cochran and Cox (1960) and standard error of difference (S.E.D.) was used for the separation of means.

The variation due to hybrids was separated into male sets, female sets and male \times female interaction sets. The mean squares of male sets, female sets and female \times male sets were subjected to *F*-test using their respective interactions with the environment. The male \times female \times environment within sets mean squares were tested using the pooled error mean squares. The main effects of male sets plus female sets constituted the general combining ability (GCA) effects and that of male \times female sets interaction was the specific combining ability (SCA) effects (Hallauer and Miranda, 1988). Similarly, ANOVA of PVA was performed for the 54 selected hybrids plus one control only for the data collected under the optimal environment at Ibadan, Nigeria.

Proportionate contribution of general combining ability and specific combining ability effects of traits

The proportionate contribution of each agronomic trait was computed as percentage of the sum of squares for the crosses ascribed to GCA and SCA (Singh and Chaudhary, 1985; Annor and Badu-Apraku, 2016) and standard errors for GCA and SCA effects were estimated by the method proposed by Cox and Frey (1984). The significance of the GCA-female, GCA-male, and SCA effects of the individual inbred lines were determined using the respective standard errors. In addition, variance ratio of the mean squares of the GCA-male and female were compared (Kearsey and Pooni, 1996) to determine the relative significance of cytoplasmic effects. Similarly, the proportion of GCA and SCA effects and the significance of maternal and paternal contributions were estimated for PVA content under optimal growing conditions.

Heterotic grouping of inbred lines

The heterotic grouping based on GCA of multiple traits also referred to as the HGCAMT method (Badu-Apraku *et al.*, 2013) was employed to group the inbred lines under each and across environments. This was done by identifying the traits with significant mean squares across the test environments (low-N, drought and optimal) and standardizing their GCA effects to reduce the effects of the different rating scales used to measure the traits. Importance was, however, attached to the six traits employed in the low-N and drought base index, which included stay-green characteristic, anthesis-silking interval, plant and ear aspects, ears per plant and grain yield. Thereafter, the standardized GCA values were used for cluster analysis based on Ward's minimum variance method implemented in SAS (SAS Institute, 2012) to group the inbreds under the different environments.

Identification of inbred testers and selection of low-nitrogen and drought tolerant single crosses

The criteria proposed by Pswarayi and Vivek (2008) were adopted to identify an inbred tester. According to the criteria, a tester must (i) belong to a known heterotic group, (ii) have a high significant positive GCA across the test environments and (iii) have high yield per se. In addition to these criteria, the inbred tester was expected to have significant positive GCA-male and female effects

for PVA and its carotenoids (Halilu *et al.*, 2016). Identification of low-N and drought tolerant inbreds and hybrids was aided by the multiple trait base index (MI) under each of the stress conditions as proposed by Badu-Apraku *et al.* (2011a). A positive MI value indicated low-N or drought tolerance while a negative index denoted susceptibility of a genotype to the stressful conditions (Badu-Apraku *et al.*, 2011a). Furthermore, the MI was applied to data across drought and low-N environments to assess the reactions of the inbred lines to the two stresses.

Identification of high-yielding and stable hybrids across environments

Using the MI across drought and low-N environments, a set of 29 hybrids, including the best 15 drought and low-N tolerant, and 10 most drought and low-N susceptible hybrids, as well as the four controls were selected for G × E analysis using the GGE-biplot package (Yan, 2001). The most promising hybrids across low-N, drought and optimal environments were identified and their stability across environments was examined.

Results

Analysis of variance of agronomic traits under the contrasting environments

Across the two drought environments, the ANOVA showed significant ($P < 0.05$ or $P < 0.01$) differences among environment (E), genotype (G) and genotype × environment interaction (G × E) mean squares for grain yield and all traits except for the E mean squares for anthesis–silking interval (Table 1). Significant ($P < 0.05$ or $P < 0.01$) variations were also observed for GCA-male and female, SCA and their interactions with E for all traits except SCA × E effects for ear and plant aspects.

Across the four low-N environments, significant ($P < 0.05$ or $P < 0.01$) mean squares of E, G and G × E interaction were revealed for grain yield and other measured traits except G × E interaction for anthesis–silking interval (Table 2). The GCA-male, GCA-female, SCA, GCA-male × E, GCA-female × E, and SCA × E revealed significant ($P < 0.05$ or $P < 0.01$) differences for all traits under low-N except for SCA effect for anthesis–silking interval. Under optimal environments, significant differences were observed among E, G and G × E interaction mean squares for all traits but not for anthesis–silking interval (Table 3). Similarly, GCA-male, GCA-female and SCA effects were significant ($P < 0.05$ or $P < 0.01$) for all traits except for anthesis–silking interval. Significant ($P < 0.05$ or $P < 0.01$) GCA-male × E, GCA-female × E and SCA × E were obtained for all traits except for anthesis–silking interval. Across environments, significant ($P < 0.05$ or $P < 0.01$) differences were observed among E, G and G × E interaction mean squares for all measured traits except for anthesis–silking interval for G × E interaction (Table 3). Furthermore, effects of GCA-male, GCA-female, SCA, GCA-male × E, GCA-female × E and SCA × E were significant ($P < 0.05$ or $P < 0.01$) for all traits except for the GCA-male, GCA-female and their interactions with E for anthesis–silking interval as well as the SCA × E for anthesis–silking interval.

Analysis of variance of carotenoids and tryptophan and proportionate contributions of combining ability effects

Significant ($P < 0.05$ or $P < 0.01$) variations were detected among the hybrids for all PVA carotenoids and tryptophan (Table 4).

Also, significant ($P < 0.05$ or $P < 0.01$) differences were observed among GCA-male, GCA-female and SCA effects for all PVA carotenoids, and tryptophan except for the SCA effects of β -cryptoxanthin, β -carotene and PVA.

The contributions of GCA (male + female) sum of squares were greater than SCA effect for all traits under drought, low-N and optimal conditions as well as across environments. GCA accounted for 67.2, 58.4, 60.3 and 59.3% of the sum of squares for grain yield under drought, low-N, optimal, and across research conditions, respectively. GCA effect was 75.9% for stay-green characteristic under drought and 57.6% under low-N (Table 5). The variation among proportions of GCA-female and GCA-male effects was not significant for all traits under drought, low-N and optimal conditions with the exception of stay-green characteristic, which showed significantly greater ($P < 0.05$) GCA-female effect relative to GCA-male effect under drought, and ears per plant, which recorded significantly ($P < 0.05$) larger GCA-male effect over that of female effect under optimal conditions. The proportionate contributions of GCA-male and female effects did not significantly vary among the hybrids for all traits across environments apart from anthesis–silking interval and stay-green characteristic, which recorded significantly ($P < 0.05$) greater GCA-female effects than GCA-male effects.

The GCA effects for PVA and all carotenoids compared to their respective total genotypic sum of squares were greater than SCA effects (Fig. 1). The GCA effects accounted for 87% of the PVA, while the three PVA carotenoids (α -carotene, β -carotene and β -cryptoxanthin) had 83, 81 and 90%, respectively. The variation among GCA-male and female effects was not significant for PVA and all the carotenoids.

General combining ability effects of major agronomic traits and carotenoids of selected inbred lines

Under drought conditions, TZEIORQ 29 was the only inbred which had significant ($P < 0.05$ and $P < 0.01$) and positive GCA-male and female effects for grain yield (Table 6). Also, two inbred lines, TZEIORQ 13 and TZEIORQ 24 had significant ($P < 0.05$ and $P < 0.01$) and positive GCA-male effects for grain yield, while TZEIORQ 6 recorded significant ($P < 0.05$) and positive GCA-female effects for grain yield. In addition, TZEIORQ 13 had significant ($P < 0.05$) negative GCA-male effect for anthesis–silking interval.

Under the low-N environment, the inbreds TZEIORQ 11, TZEIORQ 59, TZEIQI 82 and TZEIORQ 2 had significant ($P < 0.05$ or $P < 0.01$) and positive GCA-male and female effects for grain yield. In contrast, TZEIORQ 13 and TZEIORQ 24 had significant ($P < 0.01$) and positive GCA-male effects for grain yield, while TZEIORQ 29, TZEIORQ 48 and TZEIORQ 43 recorded significant ($P < 0.05$ or $P < 0.01$) and positive GCA-female effects. For stay green characteristic, TZEIORQ 29 and TZEIQI 82 had significant ($P < 0.05$ or $P < 0.01$) and negative GCA-male and female effects, TZEIORQ 44 and TZEIORQ 24 had significant ($P < 0.05$) and negative GCA-male effects, while TZEIORQ 70 recorded significant ($P < 0.05$) and negative GCA-female effects (Table 6). Out of the 24 parental lines, only TZEIORQ 59 and TZEIQI 82 had significant ($P < 0.05$ or $P < 0.01$) and positive GCA-male and female effects for grain yield under optimal environments. Other inbreds that exhibited significant ($P < 0.05$ or $P < 0.01$) and positive GCA-male effects for grain yield were TZEIORQ 11, TZEIORQ 13 and TZEIORQ 2, and TZEIORQ 29 for GCA-female effects.

Table 2. Mean squares of grain yield and other agronomic traits of early maturing PVA-QPM hybrids evaluated under drought at Ikenne during the 2016/17 and 2017/18 dry seasons and under low-N conditions at Ile-Ife and Mokwa during the 2016 and 2017 growing seasons in Nigeria

Source	DF	GY	DS	ASI	PLHT	PASP	EASP	EPP	STGR
Managed drought conditions									
Env	1	278 571 020 ($P < 0.01$)	219 284 ($P < 0.01$)	4.15 (NS)	3722 ($P < 0.01$)	38.3 ($P < 0.01$)	1637 ($P < 0.01$)	3.54 ($P < 0.01$)	224 ($P < 0.01$)
Set	5	2 553 717 ($P < 0.05$)	4.7 (NS)	6.65 ($P < 0.01$)	345 ($P < 0.05$)	1.33 ($P < 0.05$)	1.85 ($P < 0.05$)	0.07 ($P < 0.05$)	1.09 ($P < 0.01$)
Env × set	5	10 288 298 (NS)	13.3 ($P < 0.01$)	0.62 (NS)	636 ($P < 0.01$)	1.39 ($P < 0.01$)	0.91 (NS)	0.08 (NS)	0.87 ($P < 0.01$)
Hybrid	99	7 678 135 ($P < 0.01$)	23.9 ($P < 0.01$)	5.05 ($P < 0.01$)	1101 ($P < 0.01$)	2.89 ($P < 0.01$)	6.78 ($P < 0.01$)	0.20 ($P < 0.01$)	1.43 ($P < 0.01$)
Male (set)	18	9 683 257 ($P < 0.01$)	23.9 ($P < 0.01$)	3.41 ($P < 0.05$)	902 ($P < 0.01$)	1.79 ($P < 0.01$)	6.27 ($P < 0.01$)	0.15 ($P < 0.01$)	0.99 ($P < 0.01$)
Female (set)	18	6 861 306 ($P < 0.01$)	12.5 ($P < 0.01$)	4.98 ($P < 0.01$)	1471 ($P < 0.01$)	2.66 ($P < 0.01$)	5.95 ($P < 0.01$)	0.19 ($P < 0.01$)	2.47 ($P < 0.01$)
Female × male (set)	54	8 068 468 ($P < 0.01$)	28.9 ($P < 0.01$)	5.45 ($P < 0.01$)	1138 ($P < 0.01$)	3.57 ($P < 0.01$)	7.88 ($P < 0.01$)	0.24 ($P < 0.01$)	1.11 ($P < 0.01$)
Hybrid × env	99	1 943 928 ($P < 0.01$)	11.6 ($P < 0.01$)	3.20 ($P < 0.01$)	265 ($P < 0.01$)	0.70 ($P < 0.05$)	1.04 ($P < 0.05$)	0.06 ($P < 0.01$)	0.91 ($P < 0.01$)
Env × male (set)	18	3 080 981 ($P < 0.01$)	14.6 ($P < 0.01$)	5.57 ($P < 0.01$)	171 ($P < 0.05$)	0.7 ($P < 0.05$)	1.60 ($P < 0.01$)	0.06 ($P < 0.05$)	1.57 ($P < 0.01$)
Env × female (set)	18	2 343 429 ($P < 0.01$)	11.3 ($P < 0.01$)	4.70 ($P < 0.01$)	346 ($P < 0.01$)	0.96 ($P < 0.05$)	1.11 ($P < 0.05$)	0.11 ($P < 0.01$)	1.31 ($P < 0.01$)
Env × female × male (set)	54	1 571 631 ($P < 0.05$)	10.8 ($P < 0.01$)	2.21 ($P < 0.05$)	247 ($P < 0.05$)	0.54 (NS)	0.75 (NS)	0.05 ($P < 0.05$)	0.54 ($P < 0.05$)
Pooled error	144	973 235	3.36	1.94	156	0.48	0.73	0.04	0.35
Low soil nitrogen conditions									
Env	3	291 853 630 ($P < 0.01$)	181.8 ($P < 0.01$)	25.5 ($P < 0.01$)	95 458 ($P < 0.01$)	35.8 ($P < 0.01$)	121 ($P < 0.01$)	895 ($P < 0.01$)	3.77 ($P < 0.01$)
Set	5	1 276 289 (NS)	10.1 ($P < 0.01$)	0.73 (NS)	1217 ($P < 0.01$)	3.29 ($P < 0.01$)	2.38 ($P < 0.01$)	6.06 (NS)	0.03 (NS)
Env × set	15	2 048 452 ($P < 0.01$)	4.2 ($P < 0.01$)	0.39 (NS)	297 ($P < 0.01$)	0.29 (NS)	1.52 ($P < 0.01$)	4.93 (NS)	0.03 (NS)
Hybrid	99	9 948 239 ($P < 0.01$)	13.1 ($P < 0.01$)	0.51 ($P < 0.05$)	1317 ($P < 0.01$)	4.83 ($P < 0.01$)	6.55 ($P < 0.01$)	0.09 ($P < 0.01$)	3.23 ($P < 0.01$)
Male (set)	18	8 473 941 ($P < 0.01$)	11.1 ($P < 0.01$)	0.59 ($P < 0.05$)	1733 ($P < 0.05$)	3.36 ($P < 0.01$)	5.72 ($P < 0.01$)	10.5 ($P < 0.01$)	0.10 ($P < 0.01$)
Female (set)	18	8 667 776 ($P < 0.01$)	11.9 ($P < 0.01$)	0.74 ($P < 0.01$)	1627 ($P < 0.01$)	4.63 ($P < 0.01$)	6.81 ($P < 0.01$)	14.7 ($P < 0.01$)	0.06 ($P < 0.01$)
Female × male (set)	54	12 215 059 ($P < 0.01$)	13.4 ($P < 0.01$)	0.42 (NS)	1174 ($P < 0.01$)	5.80 ($P < 0.01$)	7.51 ($P < 0.01$)	5.94 ($P < 0.01$)	0.11 ($P < 0.01$)
Hybrid × env	297	1 607 731 ($P < 0.01$)	2.57 ($P < 0.01$)	0.39 (NS)	254 ($P < 0.01$)	0.63 ($P < 0.01$)	0.89 ($P < 0.01$)	0.04 ($P < 0.01$)	0.95 ($P < 0.01$)
Env × male (set)	54	1 222 434 ($P < 0.05$)	2.09 ($P < 0.05$)	0.28 ($P < 0.05$)	302 ($P < 0.01$)	0.53 ($P < 0.05$)	0.8 ($P < 0.05$)	4.99 ($P < 0.05$)	0.04 ($P < 0.01$)
Env × female (set)	54	1 197 534 ($P < 0.05$)	3.23 ($P < 0.01$)	0.56 ($P < 0.05$)	326 ($P < 0.01$)	0.57 ($P < 0.05$)	0.7 ($P < 0.05$)	6.59 ($P < 0.05$)	0.03 ($P < 0.05$)
Env × female × male (set)	162	1 805 934 ($P < 0.01$)	2.29 ($P < 0.01$)	0.38 ($P < 0.05$)	218 ($P < 0.01$)	0.64 ($P < 0.05$)	0.92 ($P < 0.01$)	5.46 ($P < 0.01$)	0.04 ($P < 0.01$)
Pooled error	288	862 481	1.41	0.34	113.3	0.47	0.66	3.66	0.02

Env, environment; GY, grain yield (kg/ha); DS, days to 50% silking; ASI, anthesis-silking interval; PLHT, plant height (cm); PASP, plant aspect (1–9); EASP, ear aspect (1–9); EPP, ears per plant; STGR, stay-green characteristic (1–9).

Table 3. Mean squares of grain yield and other agronomic traits of early maturing PVA-QPM hybrids under optimal environments at Ikenne, Ile-Ife and Mokwa during the 2016 and 2017 growing seasons and across all the test environments in Nigeria

Source	DF	GY	DS	ASI	PLHT	PASP	EASP	EPP	DF	STGR
Optimal growing conditions										
Env	5	187 014 730 ($P < 0.01$)	308 ($P < 0.01$)	8.32 (NS)	101 981 ($P < 0.01$)	24.9 ($P < 0.01$)	201 ($P < 0.01$)	4.04 ($P < 0.01$)	–	–
Set	5	5 812 318 ($P < 0.01$)	5.55 ($P < 0.05$)	17.5 (NS)	1112 ($P < 0.01$)	2.02 ($P < 0.01$)	4.17 ($P < 0.05$)	0.07 ($P < 0.05$)	–	–
Env × set	25	1 423 483 (NS)	4.37 ($P < 0.01$)	20.4 (NS)	596 ($P < 0.01$)	0.62 (NS)	1.95 (NS)	0.04 ($P < 0.05$)	–	–
Hybrid	99	14 416 317 ($P < 0.01$)	19.4 ($P < 0.01$)	14.6 (NS)	1505 ($P < 0.01$)	4.06 ($P < 0.01$)	8.44 ($P < 0.01$)	0.09 ($P < 0.01$)	–	–
Male (set)	18	15 538 833 ($P < 0.01$)	17.3 ($P < 0.01$)	7.85 (NS)	2676 ($P < 0.01$)	6.57 ($P < 0.01$)	11.3 ($P < 0.01$)	0.14 ($P < 0.01$)	–	–
Female (set)	18	10 998 389 ($P < 0.01$)	21.3 ($P < 0.01$)	14.7 (NS)	1613 ($P < 0.01$)	3.18 ($P < 0.01$)	6.39 ($P < 0.01$)	0.06 ($P < 0.01$)	–	–
Female × male (set)	54	17 538 940 ($P < 0.01$)	20.8 ($P < 0.01$)	17.3 (NS)	1234 ($P < 0.01$)	4.45 ($P < 0.01$)	9.71 ($P < 0.01$)	0.10 ($P < 0.01$)	–	–
Hybrid × env	495	3 190 671 ($P < 0.01$)	4.23 ($P < 0.01$)	12.8 (NS)	504 ($P < 0.01$)	0.91 ($P < 0.01$)	2.60 ($P < 0.01$)	0.05 ($P < 0.01$)	–	–
Env × male (set)	90	2 533 737 ($P < 0.01$)	4.68 ($P < 0.01$)	9.72 (NS)	471 ($P < 0.01$)	0.87 ($P < 0.05$)	2.20 ($P < 0.05$)	0.04 ($P < 0.01$)	–	–
Env × female (set)	90	3 065 651 ($P < 0.01$)	3.11 ($P < 0.05$)	12.6 (NS)	656 ($P < 0.01$)	0.81 ($P < 0.05$)	2.46 ($P < 0.01$)	0.04 ($P < 0.01$)	–	–
Env × female × male (set)	270	3 685 994 ($P < 0.01$)	4.54 ($P < 0.01$)	14.4 (NS)	473 ($P < 0.01$)	1.01 ($P < 0.01$)	2.93 ($P < 0.01$)	0.05 ($P < 0.01$)	–	–
Pooled error	432	1 198 307	2.18	13.9	235	0.56	1.65	0.02	–	–
Across test environments										
Env	11	335 016 271 ($P < 0.01$)	2992 ($P < 0.01$)	112 ($P < 0.01$)	80 140 ($P < 0.01$)	28.4 ($P < 0.01$)	150 ($P < 0.01$)	3.66 ($P < 0.01$)	5	168 ($P < 0.01$)
Set	5	5 437 386 ($P < 0.01$)	15.7 ($P < 0.01$)	8.6 (NS)	2070 ($P < 0.01$)	5.66 ($P < 0.01$)	6.78 ($P < 0.01$)	0.12 ($P < 0.01$)	5	1.64 ($P < 0.01$)
Env × set	55	1 696 784 ($P < 0.01$)	4.99 ($P < 0.05$)	10.6 ($P < 0.05$)	476 ($P < 0.01$)	0.70 ($P < 0.05$)	1.69 ($P < 0.05$)	0.04 ($P < 0.01$)	25	1.02 ($P < 0.01$)
Hybrid	99	28 365 062 ($P < 0.01$)	41.4 ($P < 0.01$)	9.59 ($P < 0.01$)	3291 ($P < 0.01$)	10.5 ($P < 0.01$)	19.1 ($P < 0.01$)	0.26 ($P < 0.01$)	99	3.51 ($P < 0.01$)
Male (set)	18	28 550 719 ($P < 0.01$)	32.6 ($P < 0.01$)	4.75 (NS)	4481 ($P < 0.01$)	10.3 ($P < 0.01$)	18.7 ($P < 0.01$)	0.27 ($P < 0.01$)	18	1.80 ($P < 0.01$)
Female (set)	18	22 408 812 ($P < 0.01$)	37.0 ($P < 0.01$)	10.1 (NS)	4054 ($P < 0.01$)	9.05 ($P < 0.01$)	16.5 ($P < 0.01$)	0.19 ($P < 0.01$)	18	3.28 ($P < 0.01$)
Female × male (set)	54	34 936 198 ($P < 0.01$)	47.1 ($P < 0.01$)	11.2 ($P < 0.05$)	2993 ($P < 0.01$)	12.7 ($P < 0.01$)	23.3 ($P < 0.01$)	0.30 ($P < 0.01$)	54	4.19 ($P < 0.01$)
Hybrid × env	1089	2 432 847 ($P < 0.01$)	5.98 ($P < 0.01$)	7.33 (NS)	376 ($P < 0.01$)	0.82 ($P < 0.01$)	1.82 ($P < 0.01$)	0.05 ($P < 0.01$)	972	0.98 ($P < 0.01$)
Env × male (set)	198	2 278 991 ($P < 0.01$)	6.72 ($P < 0.01$)	5.89 (NS)	369 ($P < 0.01$)	0.64 ($P < 0.05$)	1.72 ($P < 0.01$)	0.04 ($P < 0.01$)	90	1.05 ($P < 0.01$)
Env × female (set)	198	2 356 833 ($P < 0.01$)	5.64 ($P < 0.01$)	7.5 (NS)	480 ($P < 0.01$)	0.81 ($P < 0.01$)	1.77 ($P < 0.01$)	0.05 ($P < 0.01$)	90	1.26 ($P < 0.01$)
Env × female × male (set)	594	2 620 762 ($P < 0.01$)	6.00 ($P < 0.01$)	8.03 (NS)	350 ($P < 0.01$)	0.86 ($P < 0.01$)	1.90 ($P < 0.01$)	0.05 ($P < 0.01$)	270	0.83 ($P < 0.01$)
Pooled error	864	1 048 844	3.41	7.45	181	0.51	1.16	0.03	432	0.44

Env, environment; GY, grain yield (kg/ha); DS, days to 50% silking; ASI, anthesis–silking interval; PLHT, plant height (cm); PASP, plant aspect (1–9); EASP, ear aspect (1–9); EPP, ears per plant; STGR, stay-green characteristic (1–9).

Table 4. Mean squares and narrow sense heritability estimates of carotenoids and tryptophan of selected early maturing PVA-QPM hybrids

Source	DF	Mean squares of carotenoids ($\mu\text{g/g DW}$)							
		Lut	Zeax	β -cryp	α -caro	β -caro	PVA	T caro	Tryp
Set	5	78.1 ($P < 0.01$)	94.4 ($P < 0.01$)	1.8 ($P < 0.05$)	0.2 ($P < 0.01$)	6.5 ($P < 0.01$)	9.7 ($P < 0.01$)	383.8 ($P < 0.01$)	0.0009 ($P < 0.01$)
Rep (set)	6	26.4 (NS)	30.1 (NS)	2.5 (NS)	0.1 (NS)	10.1 (NS)	11.3 (NS)	151.6 ($P < 0.05$)	0.0001 ($P < 0.01$)
Hybrid	54	52.7 ($P < 0.01$)	23.6 ($P < 0.01$)	1.5 ($P < 0.05$)	0.2 ($P < 0.01$)	2.2 ($P < 0.05$)	4.1 ($P < 0.01$)	95.0 ($P < 0.01$)	0.0011 ($P < 0.01$)
Male (set)	12	71.2 ($P < 0.01$)	22.6 ($P < 0.01$)	2.0 ($P < 0.01$)	0.2 ($P < 0.01$)	2.5 ($P < 0.01$)	5.2 ($P < 0.01$)	117.3 ($P < 0.01$)	0.0009 ($P < 0.01$)
Female (set)	12	87.0 ($P < 0.01$)	27.9 ($P < 0.01$)	1.7 ($P < 0.05$)	0.2 ($P < 0.01$)	3.7 ($P < 0.01$)	6.3 ($P < 0.01$)	77.0 ($P < 0.01$)	0.0012 ($P < 0.01$)
Female \times male (set)	24	21.5 ($P < 0.01$)	7.3 ($P < 0.01$)	0.4 (NS)	0.1 ($P < 0.01$)	0.3 (NS)	0.8 (NS)	36.0 ($P < 0.01$)	0.0009 ($P < 0.01$)
Error	48	2.07	2.01	0.56	0.01	0.42	0.53	7.79	0.00002

Carotenoids are abbreviated as Lut, lutein; Zeax, zeaxanthin; β -cryp, β -cryptoxanthin; α -caro, α -carotene; β -caro, β -carotene; PVA, provitamin A; T caro, total carotenoid; Tryp, tryptophan.

Across the 12 test environments, five inbred lines were good combiners for grain yield both as males and females as evident in the significant ($P < 0.05$ or $P < 0.01$) and positive GCA-male and female effects recorded for the trait. Inbred lines TZEIORQ 13 and TZEIORQ 24 were good combiners for grain yield as male parents whereas TZEIORQ 48, TZEIORQ 43 and TZEIORQ 70 were good combiners for grain yield when used as female parents (Table 6).

Assessment of the combining ability effects on carotenoids under optimal growing conditions revealed TZEIORQ 29 as the only inbred with highly significant ($P < 0.01$) positive GCA-male and GCA-female effects for PVA, α -carotene, β -carotene and β -cryptoxanthin (Table 7). Additionally, TZEIORQ 29 had significant ($P < 0.05$ and $P < 0.01$) and negative GCA-male and female effects for zeaxanthin which is a non-PVA carotenoid. Also, TZEIORQ 13 had significant ($P < 0.05$) and positive GCA-female effects for PVA and β -carotene. However, significant ($P < 0.05$) and negative GCA-male and female effects for total carotenoids were detected for TZEIORQ 13 (Table 7).

Heterotic groups and inbred testers identified across test environments

The dendrogram constructed for data across the 12 test environments using the HGCAMT method placed the parental lines into four heterotic groups when 50% of the variation among inbreds was explained (Fig. 2). Eight inbred lines TZEIORQ 69, TZEIORQ 6, TZEIORQ 15, TZEIORQ 43, TZEIORQ 47, TZEIORQ 42, TZEIORQ 40 and TZEIORQ 5 constituted heterotic group I while two inbred lines TZEIORQ 7 and TZEIORQ 41 were classified into heterotic group II. It is striking that the five inbred lines, TZEIORQ 29, TZEIORQ 11, TZEIORQ 59, TZEIORQ 82 and TZEIORQ 2 identified as good combiners for grain yield when used either as male or female parents were placed in the same heterotic group (group IV) while inbred lines TZEIORQ 13 and TZEIORQ 24 with highly significant ($P < 0.01$) positive GCA-male effects for grain yield were placed in heterotic group III across environments.

Five inbred lines, TZEIORQ 29, TZEIORQ 11, TZEIORQ 59, TZEIORQ 82 and TZEIORQ 2 had significant ($P < 0.05$ or $P < 0.01$) and positive GCA-male and female effects for grain yield across environments. In addition, the five inbred lines had moderately high to high grain yield across environments. The HGCAMT method classified all the five inbred lines into heterotic group IV (Fig. 2). Based on the combining ability effects of the five inbred lines for carotenoids accumulation, only TZEIORQ 29 obtained highly significant ($P < 0.01$) and positive GCA-male and GCA-female effects for PVA, β -carotene, α -carotene and β -cryptoxanthin. Therefore, TZEIORQ 29 was identified as the best male and female early maturing PVA-QPM inbred tester for heterotic group IV. No other inbred met the set criteria for the remaining three heterotic groups across the test environments.

Grain yield in contrasting environments, and levels of carotenoids and tryptophan under optimal conditions

Under drought environments, grain yield of the highest yielding PVA-QPM hybrid, TZEIORQ 29 \times TZEIORQ 24 significantly ($P < 0.01$) exceeded the highest yielding drought tolerant commercial hybrid control, TZEI 124 \times TZEI 25, by 31% (Table 8). Comparison of the grain yield of hybrids under drought to that of optimal environments revealed a wide yield reduction (5–

Table 5. Proportion of total genotypic sum of squares of grain yield and other agronomic traits of early maturing PVA-QPM inbreds attributable to GCA and SCA effects under drought, low-N, optimal and across environments

Trait	Managed drought			Low-N conditions			Optimal conditions			Across environments		
	GCA-male	GCA-female	SCA	GCA-male	GCA-female	SCA	GCA-male	GCA-female	SCA	GCA-male	GCA-female	SCA
Grain yield	39.3	27.9	32.8	28.9	29.5	41.6	35.3	25.0	39.8	33.2	26.1	40.7
Days to 50% anthesis	39.0	20.6	40.4	28.3	29.8	41.9	32.5	40.6	26.9	28.9	35.8	35.3
Days to 50% silking	36.6	19.1	44.3	30.6	32.6	36.8	29.1	35.9	35.0	27.9	31.7	40.4
Anthesis-silking interval	24.6	36.0	39.4	33.9	42.0	24.1	19.7	36.8	43.5	18.2	38.7	43.1
Plant height	25.7	41.9	32.4	38.2	35.9	25.9	48.5	29.2	22.3	38.9	35.2	26.0
Ear height	24.0	40.9	35.2	40.1	33.9	26.0	48.1	33.7	18.2	40.1	36.2	23.7
Plant aspect	22.4	33.2	44.5	24.4	33.6	42.1	46.2	22.4	31.4	32.1	28.2	39.7
Ear aspect	31.2	29.6	39.2	28.5	34.0	37.5	41.2	23.3	35.5	32.0	28.2	39.8
Ears per plant	25.3	32.8	41.9	38.7	24.2	37.2	48.8	18.9	32.4	35.0	25.1	39.9
Stay-green characteristic	21.8	54.1	24.1	22.7	34.9	42.4	-	-	-	19.4	35.4	45.2

GCA-male, general combining ability effects of inbreds as male parents; GCA-female, general combining ability effects of inbreds as female parents; SCA, specific combining ability effects of inbreds

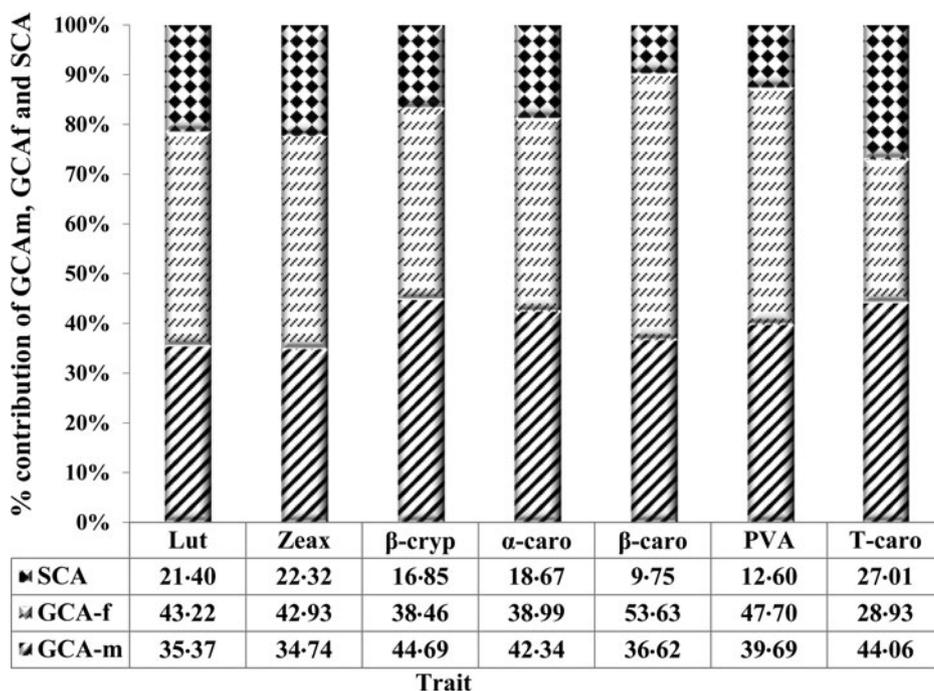


Fig. 1. Proportion of total genotypic sum of squares for carotenoids of early maturing PVA-QPM inbreds attributable to GCA of male and female (GCA-m and GCA-f) and SCA effects. Carotenoids are abbreviated as Lut, lutein; Zeax, zeaxanthin; β-cryp, β-cryptoxanthin; α-car, alpha-carotene; β-car, β-carotene; PVA, provitamin A; T-car, total carotenoids.

84%) with a mean of 37%. Under low-N conditions, the highest-ranking hybrid, TZEIORQ 29 × TZEIORQ 43 significantly ($P < 0.05$) out-yielded the best PVA hybrid control, TZEIOR 127 × TZEIOR 57, by 19%. Comparison of the yield under low-N and that under optimal environments revealed yield reduction varying from 7% for TZEIORQ 29 × TZEIORQ 43 to 80% for TZEIORQ 45 × TZEIORQ 43 with a mean of 33%. The low-N tolerant hybrids recorded lower grain yield reductions than the susceptible hybrids. The performance of the

hybrids under optimal conditions revealed the commercial normal yellow endosperm hybrid control, TZEI 124 × TZEI 25 as comparable in yield to the top performing PVA-QPM hybrids including TZEIORQ 40 × TZEIORQ 26, TZEIORQ 23 × TZEIORQ 44, TZEIORQ 26 × TZEIORQ 47 and TZEIORQ 29 × TZEIORQ 44. Across drought, low-N and optimal environments, grain yield of the best performing hybrid (TZEIORQ 40 × TZEIORQ 26) according to the multiple trait base index for combined drought and low-N tolerance was not significantly different from that of the

Table 6. GCA effects of grain yield of early PVA-QPM inbreds evaluated under drought, low-N, optimal and across environments as well as ASI and stay-green characteristic under stress for one years in Nigeria

Inbred	Managed drought conditions				Low-N conditions				Optimal conditions		Across environments			kg/ha
	Grain yield		ASI		Grain yield		STGR		Grain yield		Grain yield			
	GCA-m	GCA-f	GCA-m	GCA-f	GCA-m	GCA-f	GCA-m	GCA-f	GCA-m	GCA-f	GCA-m	GCA-f		
TZEIORQ 69	-501 (NS)	-891 (<i>P</i> <0.05)	0.2 (NS)	0.1 (NS)	25.0 (NS)	-326 (NS)	0.3 (NS)	-0.1 (NS)	68.3 (NS)	49.9 (NS)	-42.2 (NS)	231 (NS)	1174	
TZEIORQ 29	867 (<i>P</i> <0.05)	1508 (<i>P</i> <0.01)	-0.4 (NS)	-0.7 (NS)	280 (NS)	1041 (<i>P</i> <0.01)	-0.3 (<i>P</i> <0.05)	-0.6 (<i>P</i> <0.01)	199 (NS)	648 (<i>P</i> <0.05)	338 (<i>P</i> <0.05)	921 (<i>P</i> <0.01)	727	
TZEIORQ 45	647 (NS)	-1054 (<i>P</i> <0.01)	0.3 (NS)	1.4 (<i>P</i> <0.05)	-129 (NS)	-1148 (<i>P</i> <0.01)	-0.1 (NS)	0.7 (<i>P</i> <0.01)	-208 (NS)	-1091 (<i>P</i> <0.01)	-38.6 (NS)	-1104 (<i>P</i> <0.01)	1036	
TZEIORQ 48	-1013 (<i>P</i> <0.05)	437 (NS)	-0.1 (NS)	-0.7 (NS)	-176 (NS)	433 (<i>P</i> <0.05)	0.1 (NS)	-0.1 (NS)	-59.0 (NS)	393 (NS)	-257 (NS)	414 (<i>P</i> <0.01)	1385	
TZEIORQ 11	268 (NS)	58.6 (NS)	0.2 (NS)	-0.0 (NS)	411 (<i>P</i> <0.05)	559 (<i>P</i> <0.01)	0.1 (NS)	-0.2 (NS)	453(<i>P</i> <0.05)	270 (NS)	407 (<i>P</i> <0.01)	331 (<i>P</i> <0.05)	1082	
TZEIORQ 20	77.0 (NS)	67.8 (NS)	0.0 (NS)	-0.1 (NS)	-25.9 (NS)	34.8 (NS)	0.1 (NS)	0.2 (NS)	-271 (NS)	372 (NS)	-130 (NS)	209 (NS)	662	
TZEIORQ 6	-79.3 (NS)	865 (<i>P</i> <0.05)	-0.1 (NS)	-0.4 (NS)	-397 (<i>P</i> <0.05)	-46.7 (NS)	0.2 (NS)	-0.1 (NS)	-205 (NS)	-11.8 (NS)	-248 (NS)	122 (NS)	935	
TZEIORQ 44	-266 (NS)	-991 (<i>P</i> <0.05)	-0.1 (NS)	0.5 (NS)	12.0 (NS)	-547 (<i>P</i> <0.01)	-0.3 (<i>P</i> <0.05)	0.2 (NS)	22.9 (NS)	-630 (<i>P</i> <0.05)	-28.8 (NS)	-662 (<i>P</i> <0.01)	932	
TZEIORQ 42	353 (NS)	260 (NS)	0.6 (NS)	0.0 (NS)	-98.2 (NS)	148 (NS)	0.1 (NS)	-0.1 (NS)	-144 (NS)	-388 (NS)	-45.3 (NS)	-103 (NS)	814	
TZEIORQ 59	269 (NS)	323 (NS)	-0.5 (NS)	0.1 (NS)	340 (<i>P</i> <0.05)	463 (<i>P</i> <0.05)	-0.2 (NS)	-0.3 (NS)	684 (<i>P</i> <0.01)	12.3 (<i>P</i> <0.01)	501 (<i>P</i> <0.01)	714 (<i>P</i> <0.01)	1065	
TZEIORQ 15	-634 (NS)	1.7 (NS)	0.2 (NS)	-0.4 (NS)	-139 (NS)	-310 (NS)	-0.1 (NS)	-0.2 (NS)	-312 (NS)	35.6 (NS)	-308 (<i>P</i> <0.05)	-83.9 (NS)	1228	
TZEIORQ 23	11.4 (NS)	-585 (NS)	-0.3 (NS)	0.3 (NS)	-103 (NS)	-301 (NS)	0.1 (NS)	0.5 (<i>P</i> <0.01)	-229 (NS)	-660 (<i>P</i> <0.01)	-148 (NS)	-527 (<i>P</i> <0.01)	837	
TZEIQ 82	221 (NS)	27.2 (NS)	0.3 (NS)	0.4 (NS)	806 (<i>P</i> <0.01)	488 (<i>P</i> <0.05)	-0.4 (<i>P</i> <0.05)	-0.4 (<i>P</i> <0.05)	825 (<i>P</i> <0.01)	482 (<i>P</i> <0.05)	719 (<i>P</i> <0.01)	410 (<i>P</i> <0.01)	858	
TZEIORQ 47	430 (NS)	-472 (NS)	0.2 (NS)	0.8 (NS)	38.5 (NS)	-108 (NS)	-0.2 (NS)	0.1 (NS)	1278 (NS)	-149 (NS)	149 (NS)	-189 (NS)	1097	
TZEIORQ 7	-1733 (<i>P</i> <0.01)	297 (NS)	0.7 (NS)	-0.8 (NS)	-1479 (<i>P</i> <0.01)	-180 (NS)	0.7 (<i>P</i> <0.01)	0.4 (<i>P</i> <0.05)	-1514 (<i>P</i> <0.01)	-133 (NS)	-1541 (<i>P</i> <0.01)	-79.1 (NS)	992	
TZEIORQ 13	1082 (<i>P</i> <0.05)	148 (NS)	-1.2 (<i>P</i> <0.05)	-0.3 (NS)	634 (<i>P</i> <0.01)	-200 (NS)	-0.2 (NS)	0.1 (NS)	561 (<i>P</i> <0.05)	-199 (NS)	673 (<i>P</i> <0.01)	-142 (NS)	1220	
TZEIORQ 2	383 (NS)	47.6 (NS)	-0.2 (NS)	-0.8 (NS)	840 (<i>P</i> <0.01)	584 (<i>P</i> <0.01)	-0.2 (NS)	0.0 (NS)	1061 (<i>P</i> <0.01)	379 (NS)	927 (<i>P</i> <0.01)	396 (<i>P</i> <0.05)	669	
TZEIORQ 5	448 (NS)	46.9 (NS)	-0.2 (NS)	-0.0 (NS)	95.8 (NS)	-235 (NS)	-0.1 (NS)	-0.1 (NS)	-246 (NS)	-110 (NS)	-35.5 (NS)	-125 (NS)	956	
TZEIORQ 26	267 (NS)	572 (NS)	-0.3 (NS)	0.1 (NS)	125 (NS)	-20.4 (NS)	0.1 (NS)	0.1 (NS)	172 (NS)	-0.1 (NS)	155 (NS)	83.3 (NS)	958	
TZEIORQ 41	-1098 (<i>P</i> <0.05)	-667 (<i>P</i> <0.05)	0.7 (NS)	0.8 (NS)	-1061 (<i>P</i> <0.01)	-328 (NS)	0.3 (NS)	-0.0 (NS)	-987 (<i>P</i> <0.01)	-269 (NS)	-1046 (<i>P</i> <0.01)	-354 (<i>P</i> <0.05)	801	
TZEIORQ 24	1574. <i>P</i> <0.01)	-1045 (<i>P</i> <0.01)	-0.4 (NS)	0.1 (NS)	581 (<i>P</i> <0.01)	-881 (<i>P</i> <0.01)	-0.3 (<i>P</i> <0.05)	0.5 (<i>P</i> <0.01)	303 (NS)	-463 (<i>P</i> <0.05)	607 (<i>P</i> <0.01)	-718 (<i>P</i> <0.01)	454	
TZEIORQ 43	-537 (NS)	282 (NS)	0.2 (NS)	0.5 (NS)	35.4 (NS)	385 (<i>P</i> <0.05)	0.4 (<i>P</i> <0.05)	-0.2 (NS)	-567 (<i>P</i> <0.05)	251 (NS)	-361 (<i>P</i> <0.05)	283 (<i>P</i> <0.05)	872	
TZEIORQ 40	-512 (NS)	456 (NS)	-0.1 (NS)	-0.2 (NS)	-403 (<i>P</i> <0.05)	324 (NS)	-0.2 (NS)	0.1 (NS)	-123 (NS)	-95.4 (NS)	-281 (<i>P</i> <0.05)	119 (NS)	1068	
TZEIORQ 70	-525 (NS)	308 (NS)	0.4 (NS)	-0.4 (NS)	-213 (NS)	171 (NS)	0.1 (NS)	-0.4 (<i>P</i> <0.05)	388 (NS)	307 (NS)	35.1 (NS)	316(<i>P</i> <0.05)	1159	
s.e.d.	380.0	331.4	0.51	0.48	169.3	167.5	0.14	0.15	199.0	218.9	133.4	135.7		

ASI, anthesis-silking interval; STGR, stay-green characteristic (1-9); GCA-m, general combining ability effects of inbred as a male parent; GCA-f, general combining ability effects of inbred as a female parent.

Table 7. GCA effects of carotenoids ($\mu\text{g/g DW}$) for 24 selected early PVA-QPM inbred lines under optimal conditions in 2018, Nigeria

Inbred	Lutein		Zeaxanthin		β -cryptoxanthin		α -carotene		β -carotene		PVA		Total carotenoids	
	GCA-m	GCA-f	GCA-m	GCA-f	GCA-m	GCA-f	GCA-m	GCA-f	GCA-m	GCA-f	GCA-m	GCA-f	GCA-m	GCA-f
TZEIORQ 29	2.4 (NS)	5.1 (NS)	-3.5 ($P < 0.05$)	-4.1 ($P < 0.01$)	1.4 ($P < 0.01$)	1.2 ($P < 0.01$)	0.5 ($P < 0.01$)	0.4 ($P < 0.01$)	1.3 ($P < 0.01$)	1.5 ($P < 0.01$)	2.3 ($P < 0.01$)	2.3 ($P < 0.01$)	2.0 (NS)	4.1 (NS)
TZEIORQ 45	1.0 (NS)	-2.3 (NS)	2.8 ($P < 0.05$)	0.8 (NS)	-0.5 (NS)	-0.8 (NS)	-0.1 (NS)	-0.2 (NS)	-0.2 (NS)	-0.6 (NS)	-0.5 (NS)	-1.1 (NS)	3.1 (NS)	-3.1 (NS)
TZEIORQ 48	-3.5 (NS)	-2.8 (NS)	0.7 (NS)	3.3 ($P < 0.05$)	-0.9 ($P < 0.05$)	-0.4 (NS)	-0.4 ($P < 0.05$)	-0.2 (NS)	-1.2 ($P < 0.05$)	-0.9 (NS)	-1.8 ($P < 0.05$)	-1.2 (NS)	-5.1 (NS)	-0.9 (NS)
TZEIORQ 20	-2.5 (NS)	-3.2 (NS)	1.5 (NS)	2.4 (NS)	0.5 (NS)	0.4 (NS)	0.1 (NS)	0.2 (NS)	0.4 (NS)	0.1 (NS)	0.6 (NS)	0.3 (NS)	-0.2 (NS)	-0.4 (NS)
TZEIORQ 6	-1.6 (NS)	-2.6 (NS)	-1.2 (NS)	-1.4 (NS)	-0.2 (NS)	0.3 (NS)	-0.1 (NS)	-0.1 (NS)	-0.3 (NS)	-0.3 (NS)	-0.5 (NS)	-0.2 (NS)	-3.4 (NS)	-4.0 (NS)
TZEIORQ 44	4.1 (NS)	5.8 ($P < 0.05$)	-0.3 (NS)	-1.0 (NS)	-0.3 (NS)	-0.7 (NS)	0.1 (NS)	-0.1 (NS)	-0.1 (NS)	0.2 (NS)	-0.2 (NS)	-0.1 (NS)	3.6 (NS)	4.4 (NS)
TZEIORQ 42	1.8 (NS)	0.5 (NS)	0.7 (NS)	-0.8 (NS)	0.1 (NS)	-0.1 (NS)	-0.1 (NS)	-0.2 (NS)	0.4 (NS)	0.1 (NS)	0.4 (NS)	0.1 (NS)	2.8 (NS)	-0.4 (NS)
TZEIORQ 59	2.9 (NS)	3.1 (NS)	0.6 (NS)	0.8 (NS)	-0.3 (NS)	-0.2 (NS)	0.1 (NS)	0.2 (NS)	0.1 (NS)	0.4 (NS)	-0.1 (NS)	0.4 (NS)	3.2 (NS)	4.1 (NS)
TZEIORQ 23	-4.6 (NS)	-3.6 (NS)	-1.3 (NS)	0.1 (NS)	0.3 (NS)	0.2 (NS)	0.2 (NS)	0.1 (NS)	-0.4 (NS)	-0.5 (NS)	-0.3 (NS)	-0.4 (NS)	-6.0 ($P < 0.05$)	-3.8 (NS)
TZEIORQ 47	3.9 (NS)	5.0 (NS)	2.4 (NS)	0.3 (NS)	0.1 (NS)	-0.4 (NS)	0.1 (NS)	0.1 (NS)	-0.1 (NS)	-0.9 (NS)	-0.1 (NS)	-1.1 (NS)	6.3 ($P < 0.05$)	4.0 (NS)
TZEIORQ 7	0.3 (NS)	-0.1 (NS)	0.6 (NS)	1.7 (NS)	-0.1 (NS)	0.4 (NS)	0.1 (NS)	-0.3 (NS)	-0.7 (NS)	-0.4 (NS)	-0.7 (NS)	-0.3 (NS)	0.2 (NS)	1.3 (NS)
TZEIORQ 13	-4.2 (NS)	-4.9 (NS)	-3.0 ($P < 0.05$)	-2.0 (NS)	-0.1 (NS)	0.1 (NS)	-0.1 (NS)	0.2 (NS)	0.8 (NS)	1.3 ($P < 0.05$)	0.7 (NS)	1.4 ($P < 0.05$)	-6.5 ($P < 0.05$)	-5.4 ($P < 0.05$)
TZEIORQ 2	1.5 (NS)	-0.5 (NS)	1.1 (NS)	1.1 (NS)	-0.1 (NS)	-0.1 (NS)	-0.2 (NS)	-0.1 (NS)	0.1 (NS)	0.2 (NS)	0.1 (NS)	0.1 (NS)	2.6 (NS)	0.6 (NS)
TZEIORQ 5	-2.7 (NS)	-0.9 (NS)	-0.9 (NS)	-1.4 (NS)	-0.1 (NS)	-0.2 (NS)	-0.1 (NS)	-0.1 (NS)	-0.2 (NS)	-0.3 (NS)	-0.3 (NS)	-0.4 (NS)	-3.9 (NS)	-2.8 (NS)
TZEIORQ 26	1.1 (NS)	1.4 (NS)	-0.3 (NS)	0.3 (NS)	0.2 (NS)	0.3 (NS)	0.2 (NS)	0.1 (NS)	0.1 (NS)	0.1 (NS)	0.2 (NS)	0.3 (NS)	1.2 (NS)	2.2 (NS)
TZEIORQ 24	-3.9 (NS)	2.6 (NS)	0.9 (NS)	-2.3 (NS)	0.6 (NS)	-0.1 (NS)	0.2 (NS)	-0.1 (NS)	-0.3 (NS)	-0.6 (NS)	0.1 (NS)	-0.7 (NS)	-2.6 (NS)	-0.5 (NS)
TZEIORQ 43	1.6 (NS)	-1.3 (NS)	-0.7 (NS)	1.2 (NS)	-0.3 (NS)	-0.1 (NS)	-0.1 (NS)	-0.0 (NS)	0.1 (NS)	0.1 (NS)	-0.2 (NS)	0.1 (NS)	0.5 (NS)	-0.1 (NS)
TZEIORQ 40	2.3 (NS)	-1.3 (NS)	-0.2 (NS)	1.1 (NS)	-0.3 (NS)	0.1 (NS)	-0.1 (NS)	0.1 (NS)	0.3 (NS)	0.5 (NS)	0.2 (NS)	0.7 (NS)	2.1 (NS)	0.6 (NS)
S.E.D.	2.52	2.78	1.29	1.44	0.40	0.36	0.13	0.12	0.43	0.52	0.62	0.68	2.96	2.40

GCA-m, GCA effects of inbred as a male parent; GCA-f, GCA effects of inbred as a female parent.

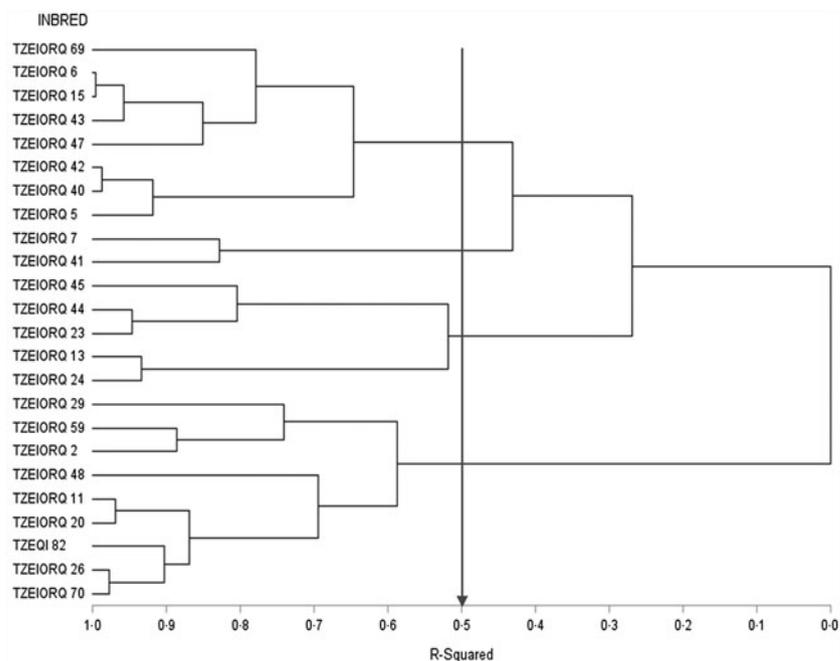


Fig. 2. Dendrogram of 24 early maturing PVA-QPM parental lines constructed from GCA effects of grain yield and other traits (HGCAMT) using Ward's minimum variance cluster analysis method across drought, low-N and optimal environments in Nigeria, 2016–17.

best commercial control (TZEI 124 × TZEI 25). Higher yield reductions were observed among susceptible hybrids across drought, low-N and optimal environments (Table 8).

Generally, the α -carotene contents of the hybrids were very low compared with the levels of the other carotenoids. PVA levels of the hybrids varied from 3.66 $\mu\text{g/g}$ for TZEIORQ 20 × TZEIORQ 48 to 9.82 $\mu\text{g/g}$ for TZEIORQ 29 × TZEIORQ 40 with a mean of 6.19 $\mu\text{g/g}$. The best-performing hybrid across test environments, TZEIORQ 40 × TZEIORQ 26, had PVA content of 6.90 $\mu\text{g/g}$. Tryptophan contents varied from 0.08% for TZEIORQ 20 × TZEIORQ 45 to 0.20% for 'Obatanpa' with a mean of 0.12%. Obatanpa was 29% higher in tryptophan content than TZEIORQ 40 × TZEIORQ 26, which was second to 'Obatanpa' in terms of tryptophan levels. The threshold of tryptophan content for a QPM genotype is 0.075% in sample in whole grain. All the hybrids had >0.075% tryptophan in sample in whole grain, suggesting that the hybrids met the quality standards of QPM genotypes. Grain yield correlated significantly and positively with PVA ($r=0.40$, $P<0.01$), α -carotene ($r=0.31$, $P<0.05$), β -cryptoxanthin ($r=0.38$, $P<0.01$) and β -carotene ($r=0.33$, $P<0.05$) but not with total carotenoids, lutein, zeaxanthin and tryptophan (Table 9).

Grain yield stability of hybrids across test environments

Due to the significant differences observed for $G \times E$ interactions for grain yield under each and across environments, the GGE biplot procedure was employed to investigate the $G \times E$ interactions and to examine yield stability across environments. The first and the second principal component axes (PC1 and PC2) showed 77.7% of the variation in grain yield (Figs 3 and 4).

The polygon view (which-won-where) of the biplot identified location-specific hybrids, with the vertex entries in a sector representing the highest-yielding genotypes in the environments that fell within the sector (Fig. 3). Thus, the vertex genotypes in each sector were more responsive to their locations than those

within the polygon and located close to the biplot origin. The polygon view displayed seven sectors with entries 2, 20, 22, 13, 19, 4 and 12 representing the vertex hybrids. Three environments, E2, E3 and E12 fell within the sector which had entry 2 (TZEIORQ 29 × TZEIORQ 43) as the vertex hybrid, implying that it was the ideal hybrid in terms of yield for those environments. Similarly, five environments E1, E4, E5, E6 and E10 were in the sector where entry 20 (TZEIORQ 26 × TZEIORQ 47) was the vertex hybrid and therefore was the highest yielding hybrid in those environments. Two vertex hybrids, entry 22 (TZEIORQ 24 × TZEIORQ 41) and entry 13 (TZEIORQ 23 × TZEIORQ 44) were placed in environment E9 and were therefore the highest-yielding hybrids in that environment. Environments E7, E8 and E11 did not have any vertex hybrids and therefore the highest yielding hybrids could not be identified in those environments. Entries 4, 12 and 19 were vertex hybrids but they were not identified with any of the environments used. The control, entry 26 (TZEIORQ 127 × TZEIORQ 57), was the least responsive genotype to environmental variability.

The 'mean performance *v.* stability' GGE biplot view was used to identify the highest yielding and most stable hybrids across the 12 test environments. In the biplot (Fig. 4), the average tester coordinate separated hybrids that had means above the grand mean of all hybrids (to the right side of the line) from those with means below the grand mean (to the left side of the line). The yield performance of the hybrids was measured by the imaginary projections from the positions of the hybrids onto the vertical axis, while stability of the hybrids was determined by their projections onto the horizontal axis. Hybrids positioned far from the vertical axis in the right direction were the higher-yielding genotypes, while those with shorter projections on to the horizontal axis were the most stable genotypes. Based on this interpretation, the entry/tester GGE biplot identified entries 22 (TZEIORQ 24 × TZEIORQ 41) and 20 (TZEIORQ 26 × TZEIORQ 47) as the highest-yielding hybrids with TZEIORQ 24 × TZEIORQ 41 as the most stable across research

Table 8. Mean grain yield performance of selected hybrids based on the multiple trait base index across environments, and levels of PVA carotenoids and tryptophan under optimal conditions in Nigeria

Hybrid	Grain yield (kg/ha)				Carotenoids ($\mu\text{g/g DW}$)				Tryptophan (%)	Yield reduction (%)		
	Drought	Low-N	Optimal	Across	β -cryp	α -caro	β -caro	PVA		Drought	Low-N	MI
TZEIORQ 40 \times TZEIORQ 26	4853	4155	5641	5014	3.32	1.30	4.60	6.90	0.14	14.0	26.3	8.09
TZEIORQ 23 \times TZEIORQ 44	3854	4415	5589	4908	2.79	1.11	3.39	5.34	0.12	31.0	21.0	7.23
TZEIORQ 47 \times TZEIORQ 23	4166	4161	5054	4608	2.64	1.04	3.04	5.11	0.12	17.6	17.7	7.14
TZEIORQ 26 \times TZEIORQ 47	5085	4160	5834	5151	2.64	1.04	3.03	4.87	0.11	12.8	28.7	6.71
TZEIORQ 42 \times TZEIORQ 20	3908	4584	5286	4823	2.95	0.36	3.94	5.60	0.14	26.1	13.3	6.67
TZEIORQ 24 \times TZEIORQ 41	4499	3950	5776	4955	2.79	1.29	5.00	7.03	0.08	22.1	31.6	6.66
TZEIORQ 20 \times TZEIORQ 45	5012	3961	5396	4854	2.98	1.01	3.90	5.90	0.08	7.12	26.6	6.57
TZEIORQ 29 \times TZEIORQ 43	2866	4694	5055	4549	4.06	1.31	7.09	9.78	0.14	43.3	7.14	6.29
TZEIORQ 48 \times TZEIORQ 43	3519	4623	5322	4812	2.55	0.82	4.05	5.73	0.10	33.9	13.1	6.05
TZEIORQ 7 \times TZEIORQ 42	4781	4016	5372	4822	3.44	0.57	4.52	6.53	0.11	11.0	25.2	6.01
TZEIORQ 43 \times TZEIORQ 5	3886	4359	5862	5031	2.68	0.96	3.61	5.43	0.11	33.7	25.6	5.87
TZEIORQ 6 \times TZEIORQ 29	4728	3322	5042	4416	4.83	1.50	5.11	8.27	0.10	6.23	34.1	5.45
TZEIORQ 29 \times TZEIORQ 24	5223	3792	5500	4642	4.94	1.46	5.64	8.84	0.13	5.04	31.1	5.29
TZEI 124 \times TZEI 25 (control 2)	3607	3690	5895	4779	–	–	–	–	–	38.8	37.4	5.19
TZEIORQ 26 \times TZEIORQ 13	4160	4384	5795	5052	2.72	0.78	4.63	6.38	0.10	28.2	24.4	5.02
TZEIOR 127 \times TZEIOR 57 (control 1)	2749	3805	5083	4268	4.95	1.44	5.26	8.46	–	45.9	25.1	4.13
TZEIORQ 29 \times TZEIORQ 40	3680	3784	5252	4500	4.19	1.44	7.01	9.82	0.14	29.9	28.0	3.97
TZEIORQ 42 \times TZEIORQ 6	3502	3462	3926	3701	2.67	0.74	3.77	5.48	0.11	10.8	11.8	2.64
TZEIORQ 7 \times TZEIORQ 59	2753	3268	5245	4171	2.35	0.59	4.05	5.52	0.10	47.5	37.7	2.14
TZE PoP DT STR \times TZEI 17 (control 4)	3147	2962	5156	4102	–	–	–	–	–	39.0	42.6	1.96
TZEIORQ 2 \times TZEIORQ 7	1494	2327	3694	2877	2.29	0.72	2.79	4.30	0.11	59.6	37.0	–1.36
TZE PoP DT STR \times TZEI 13 (control 3)	2265	2430	4052	3213	–	–	–	–	–	44.1	40.0	–2.74
TZEIORQ 20 \times TZEIORQ 48	1195	2166	4555	3199	1.82	0.57	2.47	3.66	0.12	73.8	52.4	–5.39
TZEIORQ 42 \times TZEIORQ 44	569	1163	1667	1316	1.51	0.80	3.60	4.75	0.14	65.9	30.3	–9.08
TZEIORQ 5 \times TZEIORQ 7	541	823	3419	2007	2.20	0.83	2.59	4.10	0.11	84.2	75.9	–11.3
TZEIORQ 47 \times TZEIORQ 42	617	840	2859	1729	1.67	0.78	3.95	5.18	0.14	78.4	70.6	–14.0
TZEIORQ 45 \times TZEIORQ 43	697	478	2367	1376	1.86	0.62	4.30	5.54	0.12	70.6	79.8	–15.6
OBATANPA (QPM standard control)	–	–	–	–	–	–	–	–	0.20	–	–	–
MEAN	3246	3341	4766	4032	2.95	0.96	4.22	6.19	0.12	36.5	33.1	
S.E.D.	521.7	355.5	343.5	226.1	0.6	0.062	0.867	0.931	0.004			

Carotenoids: β -cryp, β -cryptoxanthin; α -caro, alpha-carotene; β -caro, β -carotene; PVA, provitamin A; Across, across environments; MI, multiple trait base index across environments.

Table 9. Phenotypic correlation among traits of selected PVA-QPM hybrids under optimal environments in Nigeria, 2018

Trait	PVA	β -cryp	α -caro	β -caro	Tcaro	Lut	Zeax	Tryp
β -cryp	0.73 ($P < 0.0001$)							
α -caro	0.63 ($P < 0.0001$)	0.72 ($P < 0.0001$)						
β -caro	0.94 ($P < 0.0001$)	0.51 ($P < 0.0001$)	0.47 ($P < 0.01$)					
Tcaro	0.33 ($P < 0.05$)	0.29 ($P < 0.05$)	0.39 ($P < 0.01$)	0.32 ($P < 0.05$)				
Lut	0.04 (NS)	0.02 (NS)	0.21 (NS)	0.09 (NS)	0.83 ($P < 0.0001$)			
Zeax	0.15 (NS)	0.24 (NS)	0.13 (NS)	0.04 (NS)	0.54 ($P < 0.0001$)	0.14 (NS)		
Tryp	0.08 (NS)	0.06 (NS)	0.18 (NS)	0.05 (NS)	0.24 (NS)	0.12 (NS)	0.13 (NS)	
GY	0.40 ($P < 0.01$)	0.38 ($P < 0.01$)	0.31 ($P < 0.05$)	0.33 ($P < 0.05$)	0.18 (NS)	0.04 (NS)	0.21 (NS)	-0.09 (NS)

PVA, provitamin A; β -cryp, β -cryptoxanthin; α -caro, alpha-carotene; β -caro, β -carotene; Tcaro, total carotenoid; Lut, lutein; Zeax, zeaxanthin; Tryp, tryptophan; GY, grain yield across drought, low-N and optimal environments.

environments. These two hybrids were apparently among the top six high-yielding hybrids identified by the multiple trait base index for combined drought and low-N tolerance. The highest yielding and most stable hybrid, TZEIORQ 24 \times TZEIORQ 41, out-yielded the best commercial hybrid control, entry 27 (TZEI 124 \times TZEI 25), by 4% across test environments.

Discussion

The significant mean squares of hybrid and environment observed for grain yield and most of the measured traits under low-N, drought, optimal and across environments indicated the presence of genetic variation among the hybrids (Badu-Apraku and Oyekunle, 2012) and that the test environments were unique and could detect genetic differences among the hybrids. The significant mean squares of G \times E interaction observed for the important agronomic traits including grain yield under each and across test environments suggested that environmental differences influenced the expression of the measured traits. This result substantiated the need to conduct genotype evaluations across multiple environments (Najafian *et al.*, 2010; Zali *et al.*, 2011; Badu-Apraku *et al.*, 2011a, 2011b) and to employ one of the available statistical tools for assessing yield performance and stability of genotypes (Yan *et al.*, 2000). The hybrid \times research condition interaction mean squares were not significant for the measured traits including grain yield across environments, implying that there was consistency in the ranking of the hybrids across research conditions and that hybrid performance in the varying environments was influenced significantly by their genetic potential. This result also implied that selection for a hybrid that performed well across drought or low-N environments will not carry a yield penalty under optimal conditions, as reported by Bolaños and Edmeades (1993).

The significant mean squares observed for all carotenoids and tryptophan indicated high genetic variation among the hybrids to allow genetic gains from selection. This agreed with earlier reports that there was adequate genetic variability in carotenoids within yellow maize germplasm in the tropics (Menkir *et al.*, 2008; Suwarno *et al.*, 2015). The preponderance of GCA (male + female) effects over SCA for grain yield and most measured agronomic traits under the contrasting environments indicated that additive gene effect was greater than the non-additive and that GCA largely controlled the inheritance of the traits measured for the 96 early PVA-QPM hybrids. This result suggested that superior hybrids could be produced from crosses between parents with significant and positive GCA effects (Badu-Apraku *et al.*, 2013). Several authors have reported similar results under drought (Betrán *et al.*, 2003; Badu-Apraku *et al.*, 2004; Hallauer *et al.*, 2010; Adebayo *et al.*, 2014; Oyekunle and Badu-Apraku, 2014; Annor and Badu-Apraku, 2016) and under low-N (Lafitte and Edmeades, 1995; Kling *et al.*, 1997; Ifie *et al.*, 2015). Additionally, the relative importance of variances of GCA effects over the variances of SCA effects may be due to the initial subjection of the inbred lines used in the study to low-N and drought conditions, as reported by Makumbi *et al.* (2011). However, this result is not consistent with the findings of other studies where the inheritance of grain yield and most agronomic traits of maize hybrids were influenced by non-additive genetic factors under drought (Meseka *et al.*, 2013), under low-N (Betrán *et al.*, 2003; Meseka *et al.*, 2006; Makumbi *et al.*, 2011) and under optimal conditions (Fan *et al.*, 2014). The inconsistencies in the type of gene action identified by the different authors

Code	Environment
E1	Ikenne drought, 2016/2017
E2	Ile-Ife low-N, 2016
E3	Mokwa low-N 2016
E4	Ile-Ife low-N, 2017
E5	Mokwa low-N, 2017
E6	Kadawa drought, 2017/2018
E7	Ile-Ife optimal, 2016
E8	Ikenne optimal, 2016
E9	Mokwa optimal, 2016
E10	Ile-Ife optimal, 2017
E11	Ikenne optimal, 2017
E12	Mokwa optimal, 2017

Entry	Hybrid
1	TZEIORQ 29 × TZEIORQ 24
2	TZEIORQ 29 × TZEIORQ 43
3	TZEIORQ 29 × TZEIORQ 40
4	TZEIORQ 45 × TZEIORQ 43
5	TZEIORQ 48 × TZEIORQ 43
6	TZEIORQ 11 × TZEIORQ 48
7	TZEIORQ 20 × TZEIORQ 45
8	TZEIORQ 20 × TZEIORQ 48
9	TZEIORQ 6 × TZEIORQ 29
10	TZEIORQ 42 × TZEIORQ 20
11	TZEIORQ 42 × TZEIORQ 6
12	TZEIORQ 42 × TZEIORQ 44
13	TZEIORQ 23 × TZEIORQ 44
14	TZEIORQ 47 × TZEIORQ 42
15	TZEIORQ 47 × TZEIORQ 23
16	TZEIORQ 7 × TZEIORQ 42
17	TZEIORQ 7 × TZEIORQ 59
18	TZEIORQ 2 × TZEIORQ 7
19	TZEIORQ 5 × TZEIORQ 7
20	TZEIORQ 26 × TZEIORQ 47
21	TZEIORQ 26 × TZEIORQ 13
22	TZEIORQ 24 × TZEIORQ 41
23	TZEIORQ 43 × TZEIORQ 5
24	TZEIORQ 43 × TZEIORQ 41
25	TZEIORQ 40 × TZEIORQ 26
26	Control 1-TZEIOR 127 × TZEIOR 57
27	Control 2-TZEI 124 × TZEI 25
28	Control 3-TZE Pop DT STR × TZEI 13
29	Control 4-TZE Pop DT STR × TZEI 17

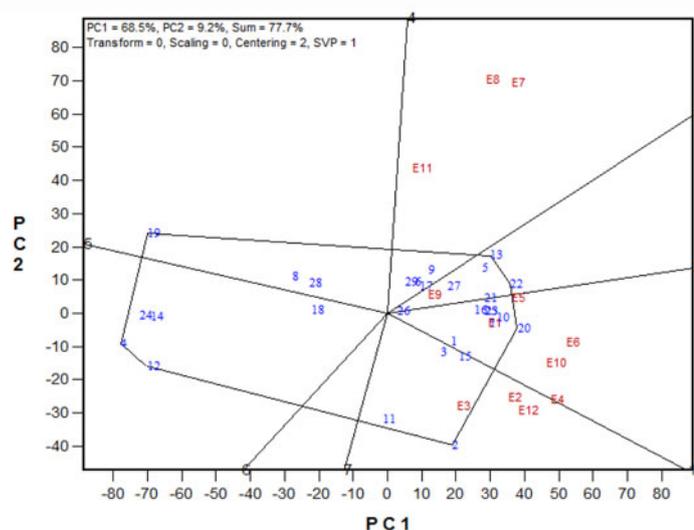


Fig. 3. (Colour online) A 'which-won-where' genotype plus $G \times E$ interaction biplot of grain yield of 25 (best 15 and worst 10 based on the combined drought and low-N base index) selected early maturing PVA-QPM hybrids plus four controls evaluated across drought, low-N and optimal environments from 2016 to 2018 in Nigeria.

may be due to the variations in the inbred lines studied, the level of inbreeding and the intensity of drought or low-N imposed. Another implication of the superiority of GCA over SCA is that recurrent selection in a population formed by inter-crossing the set of inbred lines that form the hybrids could lead to increase in the frequency of the beneficial alleles (Sprague and Tatum, 1942; Iglesias, 1989; Topal *et al.*, 2004; Chigeza *et al.*, 2014).

The significant GCA (male + female) effects for PVA and other carotenoids, as well as SCA effects for most carotenoids suggested that additive and non-additive genetic factors were equally important in the inheritance of these traits. However, the preponderance of GCA effects over SCA for the PVA and the other carotenoids indicated that GCA was the major contributor to the heritable variation in PVA carotenoids of the hybrids. This finding agreed with reports of other studies (Egesel *et al.*, 2003; Suwarno, 2012; Menkir *et al.*, 2014; Owens *et al.*, 2014; Suwarno *et al.*, 2015), which revealed that accumulation of PVA carotenoids in maize endosperm is largely influenced by additive genetic effects. This observation suggested that favourable alleles of PVA could be pyramided in the inbreds to produce outstanding hybrids and synthetics (Menkir *et al.*, 2017). The preponderance

of GCA effects compared to SCA effects indicated that early generation testing for high PVA using the set of 24 inbred lines would be effective (Fasahat *et al.*, 2016). The result also implied that the adoption of recurrent selection methods for the improvement of PVA content would be successful (Sprague and Tatum, 1942; Coors, 1999). This finding, however, contradicted the report by Halilu *et al.* (2016), who found non-additive genetic variance to be higher than the additive for all the carotenoids measured, including PVA.

The non-significant variations observed among the contributions of GCA-male and GCA-female for the traits measured including grain yield under the contrasting environments implied that maternal and paternal effects were equally important in the inheritance of the traits measured for the hybrids. This result supported the findings of Annor and Badu-Apraku (2016), who found equal contributions of the GCA-male and GCA-female sum of squares for grain yield and other measured traits under drought. Similar result was reported by Derera *et al.* (2008) and Khehra and Bhalla (1976) under optimal conditions. These results, however, are inconsistent with those of Derera *et al.* (2008) who reported that cytoplasmic effect influenced grain

Code	Environment
E1	Ikenne drought, 2016/2017
E2	Ile-Ife low-N, 2016
E3	Mokwa low-N 2016
E4	Ile-Ife low-N, 2017
E5	Mokwa low-N, 2017
E6	Kadawa drought, 2017/2018
E7	Ile-Ife optimal, 2016
E8	Ikenne optimal, 2016
E9	Mokwa optimal, 2016
E10	Ile-Ife optimal, 2017
E11	Ikenne optimal, 2017
E12	Mokwa optimal, 2017

Entry	Hybrid
1	TZEIORQ 29 × TZEIORQ 24
2	TZEIORQ 29 × TZEIORQ 43
3	TZEIORQ 29 × TZEIORQ 40
4	TZEIORQ 45 × TZEIORQ 43
5	TZEIORQ 48 × TZEIORQ 43
6	TZEIORQ 11 × TZEIORQ 48
7	TZEIORQ 20 × TZEIORQ 45
8	TZEIORQ 20 × TZEIORQ 48
9	TZEIORQ 6 × TZEIORQ 29
10	TZEIORQ 42 × TZEIORQ 20
11	TZEIORQ 42 × TZEIORQ 6
12	TZEIORQ 42 × TZEIORQ 44
13	TZEIORQ 23 × TZEIORQ 44
14	TZEIORQ 47 × TZEIORQ 42
15	TZEIORQ 47 × TZEIORQ 23
16	TZEIORQ 7 × TZEIORQ 42
17	TZEIORQ 7 × TZEIORQ 59
18	TZEIORQ 2 × TZEIORQ 7
19	TZEIORQ 5 × TZEIORQ 7
20	TZEIORQ 26 × TZEIORQ 47
21	TZEIORQ 26 × TZEIORQ 13
22	TZEIORQ 24 × TZEIORQ 41
23	TZEIORQ 43 × TZEIORQ 5
24	TZEIORQ 43 × TZEIORQ 41
25	TZEIORQ 40 × TZEIORQ 26
26	Control 1-TZEIOR 127 × TZEIOR 57
27	Control 2-TZEI 124 × TZEI 25
28	Control 3-TZE Pop DT STR × TZEI 13
29	Control 4-TZE Pop DT STR × TZEI 17

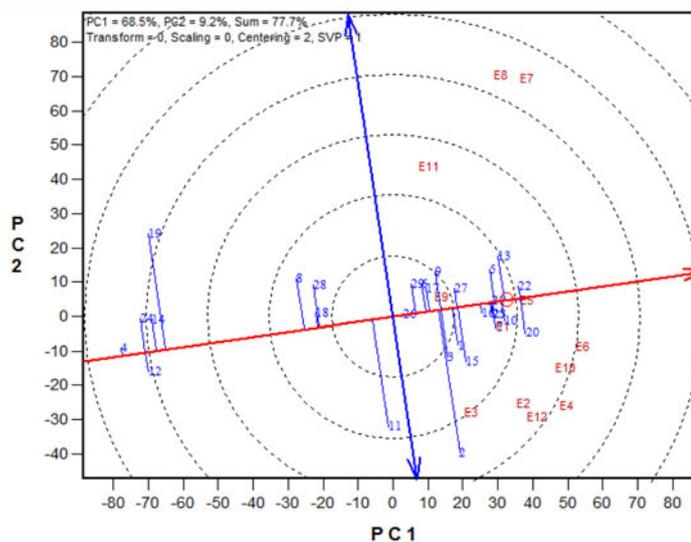


Fig. 4. (Colour online) An entry/tester genotype main effect plus $G \times E$ biplot of grain yield for 25 (best 15 and worst 10 based on the combined drought and low-N base index) selected early maturing PVA-QPM hybrids plus four controls evaluated across drought, low-N and optimal environments from 2016 to 2018 in Nigeria.

yield, prolificacy, anthesis–silking interval and ear aspect under drought, as well as anthesis–silking interval, ears per plant and ear aspect under both drought and optimal conditions. Similar inconsistent results were reported by Adebayo *et al.* (2014), who found that paternal effects modified the inheritance of ear aspect under drought. The significantly greater GCA-female effects relative to GCA-male effects recorded for stay-green characteristic under drought and across environments implied that inbred lines with significant negative GCA-female effects for stay-green characteristic should be used as females in their crosses to take maximum advantage of the effects of cytoplasmic inheritance on leaf senescence in their progenies under drought. Also, the significantly large GCA-female effects of anthesis–silking interval relative to that of GCA-male across research environments suggested that maternal effects controlled the inheritance of anthesis–silking interval in this set of inbred lines and that inbred lines with significant negative GCA-female effects for anthesis–silking interval should be used as females in their crosses to contribute to reduced anthesis and silking days in their offspring across the three contrasting environments. It was, however, surprising to find non-significant contributions of GCA-male and

GCA-female effects for PVA and its component carotenoids, implying that cytoplasmic genes did not influence the inheritance of these traits in the PVA-QPM inbred lines used.

GCA effect or additive gene effect of inbred lines for a trait is useful in determining the contributions of the parental lines to their hybrids. For example, inbred lines which have highly significant positive GCA effects for grain yield under drought or low-N have high probability of contributing favourable alleles for grain yield to the progenies in a recurrent selection programme to develop drought and/or low-N tolerant populations (Iglesias, 1989; Topal *et al.*, 2004; Chigeza *et al.*, 2014). Additionally, such inbreds could be used to improve existing populations and to develop drought and/or low-N hybrid and synthetic varieties for commercialization. The observed significant positive GCA-male and GCA-female effects for grain yield for inbred lines TZEIORQ 29, TZEIORQ 11, TZEIORQ 59, TZEIQI 82 and TZEIORQ 2 across drought, low-N and optimal environments suggested that these inbred lines would contribute favourable alleles for improved grain yield to their progenies when used as parental males or females. Similarly, the significant and positive GCA-female effects of grain yield observed for inbreds TZEIORQ

48, TZEIORQ 43 and TZEIORQ 70 across test environments indicated their high potentials to contribute favourable alleles for grain yield to their offspring as female parents. A similar inference could also be drawn for TZEIORQ 13 and TZEIORQ 24 as male parents since they displayed significant and positive GCA-male effects for grain yield across environments. In addition, the significant and negative GCA-male and female effects of stay green characteristic displayed by inbreds TZEIORQ 29, TZEIORQ 59 and TZEIQI 82 across the test environments was an indication that these inbreds, when used either as males or females, would contribute to the offspring, desirable alleles for delayed senescence, increased photosynthesis and hence increased assimilate production.

The significant and positive GCA-male and female effects observed for the inbred TZEIORQ 29 for PVA and its component carotenoids suggested that it contributed to increased levels of these carotenoids in its crosses either as a male or a female. The inbred could therefore be exploited for PVA favourable alleles in the development of superior hybrids and synthetics and for the improvement of the early maturing PVA-QPM inbred lines. The significant positive GCA-female effect recorded for TZEIORQ 13 for PVA and β -carotene indicated that it could be used as a female parent to make gains in PVA and β -carotene levels of its hybrids. However, the significant negative GCA-male and GCA-female effects for total carotenoids detected for TZEIORQ 13 implied that it could contribute to reduced total carotenoids in its hybrids when used as a female parent. This result could be a disadvantage for TZEIORQ 13 because high levels of total carotenoids could be an advantage if the influx of total carotenoids favours the accumulation of PVA carotenoids in the carotenoid biosynthetic pathway.

Classification of the newly developed early maturing PVA-QPM inbreds into appropriate heterotic groups is important for the exploitation of maximum heterosis through crossing of inbreds from opposing heterotic groups (Terror *et al.*, 1997). The four heterotic groups identified by the HGCAMT method across environments would increase the chances of developing novel and superior early maturing PVA-QPM hybrids and synthetics with combined drought and low-N tolerance for commercialization in SSA. Also, heterotic populations could be constituted by recombining inbred lines from the same heterotic group and improve the population through recurrent selection. The identification of TZEIORQ 29 as the best male and female inbred tester for heterotic group IV implied that the inbred could be used either as a male or a female parent to classify the other lines into heterotic groups and to develop high-yielding PVA-QPM hybrids and synthetics. It was, therefore, not surprising to find TZEIORQ 29 \times TZEIORQ 24 as the best hybrid under drought, and among the top performing hybrids across drought, low-N and optimal environments in Nigeria.

Assessment of the 96 early PVA-QPM single cross hybrids plus four controls under drought, low-N and optimal conditions was necessary to identify high-yielding hybrids under each and more importantly, across the three research environments. The average grain yield reduction (37%) recorded under drought at flowering was within the reported range of 40–90% (Grant *et al.*, 1989; Menkir and Akintunde, 2001; Badu-Apraku *et al.*, 2005; Derera *et al.*, 2008; Badu-Apraku *et al.*, 2011b). This result indicated that the induced drought at flowering was severe enough to effectively discriminate between the drought-tolerant and susceptible hybrids, thus facilitating the identification and selection of outstanding hybrids. Applying a selection intensity of 10%,

the drought base index which involves increased grain yield and prolificacy with short anthesis–silking interval and delayed leaf senescence, as well as excellent plant and ear aspects under drought (Badu-Apraku *et al.*, 2011a) identified the top performers of which TZEIORQ 29 \times TZEIORQ 24 was the highest yielding hybrid. TZEIORQ 29 \times TZEIORQ 24 yielded better than the best PVA commercial hybrid control (TZEI 124 \times TZEI 25) by 31%. TZEI 124 \times TZEI 25 is an orange endosperm single-cross hybrid that has been released in Ghana as ‘CSIR-Denbea’, in Mali as ‘Tamalaka’ and in Nigeria as ‘Sammaz 41’. This result indicated that TZEIORQ 29 \times TZEIORQ 24 would not only be useful in enhancing maize production and productivity in drought prone areas of SSA, but also consumption of this hybrid would help address the health disorders emanating from vitamin A deficiency and protein energy malnutrition (Bressani, 1992; Badu-Apraku and Fontem Lum, 2010; Pixley *et al.*, 2013).

Under low-N conditions, the 33% average grain yield reduction recorded was within the ranges of 10–50% and 20–50% reported by Wolfe *et al.* (1988) and Bänziger *et al.* (1999), respectively. This result indicated that the low-N condition imposed was adequate to effectively differentiate between low-N tolerant and susceptible hybrids. With a selection intensity of 10%, the low-N base index (Badu-Apraku *et al.*, 2011a) was effectively used to identify TZEIORQ 29 \times TZEIORQ 43 as the best hybrid which out-yielded the best control, TZEIOR 127 \times TZEIOR 57, by 25% under low-N conditions. The best control under low-N is a PVA non-QPM single cross hybrid identified as promising in previous studies and promoted to the regional trials of the IITA-MIP (Konaté *et al.*, 2017). This result, therefore, implied that the top-performing early PVA-QPM hybrid would be crucial in enhancing maize production under low-N environments in the region. The result has also highlighted the enormous progress made in the development of the early maturing PVA-QPM hybrids for low-N tolerance.

Combined assessment of grain yield performance using the multiple trait base index across drought, low-N and optimal environments revealed a set of ten top performing hybrids of which TZEIORQ 40 \times TZEIORQ 26 was the most outstanding. The top performers identified across environments were essentially among the best performers under each environment including optimal conditions. This implied that the best hybrids selected across the three research conditions would consistently display superior grain yield performance across seasons and in varying production environments in SSA without yield penalties. Although the grain yield performance of the top ten PVA-QPM hybrids was statistically the same as the best commercial orange endosperm hybrid control, TZEI 124 \times TZEI 25 under the contrasting environments, the identified hybrids have the extra advantage of elevated levels of PVA carotenoids as well as lysine and tryptophan.

The range of PVA concentrations (6.50–9.82 $\mu\text{g/g}$ DW) recorded for the top 15 hybrids in the current study was moderate compared with the 15 $\mu\text{g/g}$ DW target set by the Harvest-Plus challenge programme (Harjes *et al.*, 2008; Simpungwe *et al.*, 2017). The PVA concentrations observed for the hybrids in the current study corroborated with the report by Andersson *et al.* (2017), who indicated that although some progress has been made with respect to the improvement of PVA carotenoids in developed maize varieties in SSA, with over 40 varieties released, the PVA levels of these varieties range between 6 and 10 $\mu\text{g/g}$. This result suggested that there is the need to introgress

favourable PVA alleles from other sources to improve the tropically adapted early PVA-QPM inbred lines. Fortunately, apart from the early PVA-QPM inbred line TZEIORQ 55 (15.1 µg/g) identified in the current study, extra-early PVA inbred lines TZEEIOR 202 (23.98 µg/g) and TZEEIOR 205 (22.58 µg/g), with PVA levels exceeding the 15 µg/g target by 50.4 and 59.9%, respectively, have been identified in the IITA-MIP. The extra-early maturing PVA inbreds have resulted in the development of PVA hybrids TZEEIOR 197 × TZEEIOR 205 (20.1 µg/g) and TZEEIOR 202 × TZEEIOR 205 (22.7 µg/g), containing about double the amount of PVA of the commercial PVA hybrid control, TZEE-Y Pop STR C5 × TZEEI 58 (11.4 µg/g), which are in the pipeline for release in SSA (Badu-Apraku *et al.*, 2018). These new sources of PVA beneficial alleles should help to facilitate the accumulation of PVA carotenoid concentrations in the existing early PVA-QPM hybrids and to even exceed the set target (15 µg/g DW). Exceeding the PVA set target is a practically feasible approach to maximize the benefits of consumers of PVA maize because studies have revealed that the amount of PVA lost during storage, milling and preparation of different local food items could be about 70% (Mugode *et al.*, 2014; Pillay *et al.*, 2014; De Moura *et al.*, 2015) and the degree of loss widely varies among maize genotypes. The range of values of tryptophan content (>0.075 in whole grain sample) recorded for the hybrids indicated that all the hybrids met the quality standards of a QPM genotype (Vivek *et al.*, 2008; Teklewold *et al.*, 2015). The significant and positive correlation observed between PVA and grain yield indicated that improving the synthesis and accumulation of PVA and other carotenoids simultaneously may be effectively accomplished without reducing the grain yield potential and related important agronomic traits (Bouis and Welch, 2010; Menkir *et al.*, 2014). This finding contrasts with the results of Halilu *et al.* (2016), who found non-significant correlations among grain yield and measured carotenoid concentrations, and indicated that the traits can be improved independently.

An important objective in the current study was to identify high and stable yielding hybrids across drought, low-N and optimal environments for further testing and commercialization. Significant G × E interactions are advantageous when the objective is to develop location specific varieties characterized by narrow adaptation. However, it is a disadvantage when developing varieties for broad adaptation (Badu-Apraku and Fakorede, 2017). The observed significant genotype × environment interaction mean squares for grain yield and most other measured traits under drought, low-N, optimal, and across environments indicated that the expression of the traits varied with environments. This observation, therefore, warranted the use of the 'which-won-where' and the 'mean performance *v.* stability' GGE biplot views to identify hybrids with location-specific and broad adaptations, respectively. From the 'which-won-where' view of the GGE biplot, the first and the second principal component axes (PC1 and PC2) explained 77.7% of the variation in grain yield of the hybrids indicating that PC1 and PC2 sufficiently approximated the environment-centred data. Hybrids with broad adaptation would be most preferred by farmers, especially in SSA because of the significant variation in seasons and production environments. However, in a few instances, hybrids with location-specific adaptation may be necessary. Hybrid, TZEIORQ 29 × TZEIORQ 43 (entry 2) was the highest yielding in environments E2 (2016 low-N, Ile-Ife), E3 (2016 low-N, Mokwa) and E12 (2017 optimal, Mokwa). This result suggested that TZEIORQ 29 ×

TZEIORQ 43 would display superior performance in nitrogen deficient environments at Ile-Ife and Mokwa without compromising yield under optimal environments at Mokwa. Also, E2, E3 and E12 could constitute a mega-environment because they shared the same winning hybrid. However, several years of testing in these three locations would be needed to ascertain whether they are indeed mega-environments or not. Yan *et al.* (2000, 2007, 2010) found that selection of a mega-environment identified from location-groups depends on the consistency of the location groupings as well as the winning genotypes across years. Hybrid TZEIORQ 26 × TZEIORQ 47 (entry 20) was the highest yielding in environments E1 (2016 drought, Ikenne), E4 (2017 low-N, Ile-Ife), E5 (2017 low-N, Mokwa), E6 (2018 combined heat and drought, Kadawa) and E10 (2017 optimal, Ile-Ife). This result indicated that TZEIORQ 26 × TZEIORQ 47 would be high-yielding across drought and low-N environments at Ikenne, Kadawa, Ile-Ife and Mokwa with an acceptable performance under optimal conditions at Ile-Ife. What is also interesting with this result is that TZEIORQ 26 × TZEIORQ 47 would be the preferred hybrid under combined heat and drought stress at Kadawa located in the semi-arid/Sudan Savanna agro-ecology of Nigeria. Additionally, TZEIORQ 24 × TZEIORQ 41 (entry 22) and TZEIORQ 23 × TZEIORQ 44 (entry 13) were the highest-yielding hybrids only in environment E9 (2017 optimal, Mokwa), suggesting that these two hybrids were the most promising for the non-stressful environment at Mokwa. Moreover, the 'mean performance *v.* stability' GGE biplot view identified TZEIORQ 24 × TZEIORQ 41 (entry 22) and TZEIORQ 26 × TZEIORQ 47 (entry 20) as the best yielding single cross hybrids across the 12 research environments. Of the two hybrids, TZEIORQ 24 × TZEIORQ 41 was the most stable across the test environments and should be extensively tested on-farm for consistent performance and commercialization in SSA.

In conclusion, additive genetic factors were more significant than non-additive for grain yield, most of the measured agronomic traits under the contrasting environments and for PVA carotenoids under optimal conditions. The HGCAMT method revealed four heterotic groups for the inbreds based primarily on pedigree, with TZEIORQ 29 emerging as the best male and female tester for heterotic group IV. The hybrids TZEIORQ 29 × TZEIORQ 43, TZEIORQ 29 × TZEIORQ 40, TZEIORQ 29 × TZEIORQ 24 and TZEIORQ 6 × TZEIORQ 29 combined high-yielding ability with moderately high PVA levels and should be tested extensively for release and commercialization in SSA to reduce vitamin-A and protein deficiencies in the sub-region. The best-yielding and most adapted hybrid TZEIORQ 24 × TZEIORQ 41 should be further tested on-farm to confirm the consistency of performance under the contrasting environments for commercialization in SSA.

Acknowledgements. The authors are grateful to the staff of the IITA Maize Improvement Programme and the Food and Nutrition laboratory of IITA in Ibadan, Nigeria, for technical assistance. Finally, the authors are grateful to Abidemi Olutayo Talabi, Mustapha Abu Dadzie and Noudifoulé Tchala for their diverse contributions to this research.

Financial support. This research received financial support from the USAID through the West Africa Centre for Crop Improvement (WACCI), and the Bill and Melinda Gates Foundation through the DTMA/STMA Projects.

Conflict of interest. The authors hereby declare that the study was carried out without any financial and/or commercial relationship that could result in a potential conflict of interest.

Ethical standards. Not applicable.

References

- Adebayo MA, Menkir A, Blay E, Gracen V, Danquah E and Hearne S** (2014) Genetic analysis of drought tolerance in adapted × exotic crosses of maize inbred lines under managed stress conditions. *Euphytica* **196**, 261–270.
- Andersson MS, Saltzman A, Virk PS and Pfeiffer WH** (2017) Progress update: crop development and biofortified staple food crops under HarvestPlus. *African Journal of Food, Agriculture, Nutrition and Development* **2**, 11905–11935.
- Annor B and Badu-Apraku B** (2016) Gene action controlling grain yield and other agronomic traits of extra-early quality protein maize under stress and non-stress conditions. *Euphytica* **212**, 213–228.
- Badu-Apraku B and Fakorede MAB** (2017) Breeding of quality protein and provitamin A maize. In Badu-Apraku B and Fakorede MAB (eds), *Advances in Genetic Enhancement of Early and Extra-early Maize for Sub-Saharan Africa*. Cham, Switzerland: Springer, pp. 217–244.
- Badu-Apraku B and Fontem Lum AF** (2010) The pattern of grain yield response of normal and quality protein maize cultivars in stress and non-stress environments. *Agronomy Journal* **102**, 381–394.
- Badu-Apraku B and Oyekunle M** (2012) Genetic analysis of grain yield and other traits of extra-early yellow maize inbreds and hybrid performance under contrasting environments. *Field Crops Research* **129**, 99–110.
- Badu-Apraku B, Fakorede MAB, Menkir A, Kamara AY and Adam A** (2004) Effects of drought-screening methodology on genetic variances and covariances in Pool 16 DT maize population. *Journal of Agricultural Science* **142**, 445–452.
- Badu-Apraku B, Fakorede MAB, Menkir A, Kamara AY and Dapaah S** (2005) Screening maize for drought tolerance in the Guinea savanna of West and Central Africa. *Cereal Research Communications* **33**, 533–540.
- Badu-Apraku B, Fakorede MAB, Oyekunle M and Akinwale RO** (2011a) Selection of extra-early maize inbreds under low N and drought at flowering and grain-filling for hybrid production. *Maydica* **56**, 29–41.
- Badu-Apraku B, Fontem LA, Akinwale RO and Oyekunle M** (2011b) Biplot analysis of diallel crosses of early maturing tropical yellow maize inbreds in stress and nonstress environments. *Crop Science* **51**, 173–188.
- Badu-Apraku B, Akinwale RO, Franco J and Oyekunle M** (2012) Assessment of reliability of secondary traits in selecting for improved grain yield in drought and low-nitrogen environments. *Crop Science* **52**, 2050–2062.
- Badu-Apraku B, Oyekunle M, Fakorede MAB, Vroh I, Akinwale RO and Aderounmu M** (2013) Combining ability, heterotic patterns and genetic diversity of extra-early yellow inbreds under contrasting environments. *Euphytica* **192**, 413–433.
- Badu-Apraku B, Annor B, Oyekunle M, Akinwale RO, Fakorede MAB, Talabi AO, Akaogu IC, Melaku G and Fasanmade Y** (2015) Grouping of early maturing quality protein maize inbreds based on SNP markers and combining ability under multiple environments. *Field Crops Research* **183**, 169–183.
- Badu-Apraku B, Fakorede MAB, Talabi AO, Oyekunle M, Akaogu IC, Akinwale RO, Annor B, Melaku G, Fasanmade Y and Aderounmu M** (2016) Gene action and heterotic groups of early white quality protein maize inbreds under multiple stress environments. *Crop Science* **56**, 183–199.
- Badu-Apraku B, Talabi OA, Garcia-Oliveira AL and Gedil M** (2018) IITA scientists develop multiple stress tolerant maize hybrids with high levels of Pro-Vitamin A. *IITA News* 2463, 7th January 2019. Available at <http://bulletin.iita.org/index.php/2019/01/07/iita-scientists-develop-multiple-stress-tolerant-maize-hybrids-with-high-levels-of-pro-vitamin-a/> (Accessed 13 September 2019).
- Bänziger M, Edmeades GO and Lafitte HR** (1999) Selection for drought tolerance increases maize yields across a range of nitrogen levels. *Crop Science* **39**, 1035–1040.
- Betrán FJ, Beck D, Bänziger M and Edmeades GO** (2003) Genetic analysis of inbred and hybrid grain yield under stress and non-stress environments in tropical maize. *Crop Science* **43**, 807–817.
- Bolaños J and Edmeades GO** (1993) Eight cycles of selection for drought tolerance in lowland tropical maize. I. Responses in grain yield, biomass, and radiation utilization. *Field Crops Research* **31**, 233–252.
- Bouis HE and Welch RM** (2010) Biofortification: a sustainable agricultural strategy for reducing micronutrient malnutrition in the global south. *Crop Science* **50**, S20–S32.
- Bremner JM and Mulvaney CS** (1982) Nitrogen – total. In Page AL, Miller RH and Keeney DK (eds), *Methods of Soil Analysis. Part 2 – Chemical and Microbiological Properties*. Madison, WI, USA: Soil Science of America Inc., pp. 595–616.
- Bressani R** (1992) Nutritional value of high lysine maize in human. In Mertz ET (ed.), *Quality Protein Maize*. St Paul, MN, USA: American Association of Cereal Chemists, pp. 205–224.
- Chigeza G, Mashingaidze K and Shanahan P** (2014) Advanced cycle pedigree breeding in sunflower. II: combining ability for oil yield and its components. *Euphytica* **195**, 183–195.
- Cochran WG and Cox GM** (1960) *Experimental Designs*. New York, USA: John Wiley and Sons.
- Comstock RE and Robinson HF** (1948) The components of genetic variance in population of biparental progenies and their use in estimating the average degree of dominance. *Biometrics* **4**, 254–266.
- Coors JG** (1999) Selection methodologies and heterosis. In Coors JG and Pandey S (eds), *The Genetics and Exploitation of Heterosis in Crops*. Madison, WI, USA: American Society of Agronomy Inc., pp. 225–245.
- Cox DJ and Frey KJ** (1984) Combining ability and the selection of parents for interspecific oat matings. *Crop Science* **24**, 963–967.
- Crow JF and Kermicle J** (2002) Oliver Nelson and quality protein maize. *Genetics* **160**, 819–821.
- De Moura FF, Miloff A and Boy E** (2015) Retention of provitamin A carotenoids in staple crops targeted for biofortification in Africa: cassava, maize and sweet potato. *Critical Reviews in Food Science and Nutrition* **55**, 1246–1269.
- Derera J, Tongoona P, Vivek BS and Laing MD** (2008) Gene action controlling grain yield and secondary traits in southern African maize hybrids under drought and non-drought environments. *Euphytica* **162**, 411–422.
- Edmeades GO** (2013) *Progress in Achieving and Delivering Drought Tolerance in Maize – an Update*. Ithaca, NY, USA: The International Service for the Acquisition of Agri-biotech Applications (ISAAA).
- Egesel CO, Wong JC, Lambert RJ and Rocheford TR** (2003) Combining ability of maize inbreds for carotenoids tocopherols. *Crop Science* **43**, 818–823.
- Fan XM, Zhang YD, Yao WH, Bi YQ, Liu L, Chen HM and Kang MS** (2014) Reciprocal diallel crosses impact combining ability, variance estimation, and heterotic group classification. *Crop Science* **54**, 89–97.
- FAO** (2016) *FAOSTAT*. Rome, Italy: FAO (Accessed 18 February 2018).
- Fasahat P, Rajabi A, Rad JM and Derera J** (2016) Principles and utilization of combining ability in plant breeding. *Biometrics and Biostatistics International Journal* **4**, 1–22.
- Grant RF, Jackson BS, Kiniry JR and Arkin GF** (1989) Water-deficit timing effects on yield components in maize. *Agronomy Journal* **81**, 61–65.
- Halilu AD, Ado SG, Aba DA and Usman IS** (2016) Genetics of carotenoids for provitamin A biofortification in tropical-adapted maize. *The Crop Journal* **4**, 313–322.
- Hallauer AR and Miranda JB** (1988) *Quantitative Genetics in Maize Breeding*, 2nd Edn. Ames, IA, USA: Iowa State University Press.
- Hallauer AR, Carena MJ and Miranda-Filho JB** (2010) Testers and combining ability. In *Quantitative Genetics in Maize Breeding*, 2nd Edn. Handbook of Plant Breeding vol. 6. Ames, IA, USA: Iowa State University Press, pp. 383–423.
- Harjes CE, Rocheford TR, Bai L, Brutnell TP, Kandianis CB, Sowinski SG, Stapleton AE, Vallabhaneni R, Williams M, Wurtzel ET, Yan J and Buckler ES** (2008) Natural genetic variation in lycopene epsilon cyclase tapped for maize biofortification. *Science* **319**, 330–333.
- Hernández HH and Bates LS** (1969) *A Modified Method for Rapid Tryptophan Analysis of Maize*. CIMMYT Research Bulletin No. 13. Mexico, DF, Mexico: CIMMYT.

- Howe JA and Tanumihardjo SA (2006) Carotenoid-biofortified maize maintains adequate vitamin A status in Mongolian gerbils. *Journal of Nutrition* **136**, 2562–2567.
- Ifie BE, Badu-Apraku B, Gracen V and Danquah EY (2015) Genetic analysis of grain yield of IITA and CIMMYT early-maturing maize inbreds under *Striga*-infested and low-soil-nitrogen environments. *Crop Science* **55**, 610–623.
- Iglesias CA (1989) *Efficiency of S2 Recurrent Selection for Improvement of Exotic and Semi-Exotic Populations in Maize (Zea mays L.)* (Retrospective Theses and Dissertations). Paper 9199. Ames, Iowa: Iowa State University.
- Kearsey MJ and Pooni HS (1996) *The Genetical Analysis of Quantitative Traits*. London, UK: Chapman and Hall.
- Khehra AS and Bhalla SK (1976) Cytoplasmic effects on quantitative characters in maize (*Zea mays* L.). *Theoretical and Applied Genetics* **47**, 271–274.
- Kling JG, Oikeh SO, Akintoye HA, Heuberger HT and Horst WJ (1997) Potential for developing nitrogen use efficient maize for low input agricultural systems in the moist savannas of Africa. In Edmeades GO, Bänzinger M, Mickelson HR and Peña-Valdivia CB (eds), *Developing Drought- and Low-N Tolerant Maize. Proceedings of a Symposium, March 25–29, 1996*. El Batán, Mexico: CIMMYT, pp. 490–501.
- Konaté L, Badu-Apraku B and Traoré D (2017) Combining ability and heterotic grouping of early maturing provitamin A maize inbreds across *Striga* infested and optimal growing environments. *Journal of Agriculture and Environment for International Development* **111**, 141–157.
- Krivanek AF, De Groot HD, Gunaratna NS, Diallo AO and Friesen D (2007) Breeding and disseminating quality protein maize (QPM) for Africa. *African Journal of Biotechnology* **6**, 312–324.
- Lafitte HR and Edmeades GO (1995) Association between traits in tropical maize inbred lines and their hybrids under high and low soil nitrogen. *Maydica* **40**, 259–267.
- Ma Y and Nelson EO (1975) Amino acid composition and storage proteins in two new high-lysine mutants in maize. *Cereal Chemistry* **52**, 412–419.
- Makumbi D, Betrán FJ, Bänziger M and Ribaut J (2011) Combining ability, heterosis and genetic diversity in tropical maize (*Zea mays* L.) under stress and non-stress conditions. *Euphytica* **180**, 143–162.
- Menkir A and Akintunde AO (2001) Evaluation of the performance of maize hybrids, improved open pollinated and farmers' local varieties under well-watered and drought stress conditions. *Maydica* **46**, 227–238.
- Menkir A, Badu-Apraku B, Thé C and Adepoju A (2003) Evaluation of heterotic patterns of IITA's lowland white maize inbred lines. *Maydica* **48**, 161–170.
- Menkir A, Liu W, White WS, Maziya-Dixon B and Rocheford T (2008) Carotenoid diversity in tropical-adapted yellow maize inbred lines. *Food Chemistry* **109**, 521–529.
- Menkir A, Gedil M, Tanumihardjo S, Adepoju A and Bossey B (2014) Carotenoid accumulation and agronomic performance of maize hybrids involving parental combination from different marker-based groups. *Food Chemistry* **148**, 131–137.
- Menkir A, Maziya-Dixon B, Mengesha W, Rocheford T and Alamu EO (2017) Accruing genetic gain in pro-vitamin A enrichment from harnessing diverse maize germplasm. *Euphytica* **213**, 1–12.
- Meseka SK, Menkir A, Ibrahim AES and Ajala SO (2006) Genetic analysis of performance of maize inbred lines selected for tolerance to drought under low nitrogen. *Maydica* **51**, 487–495.
- Meseka SK, Menkir A, Ibrahim AES and Ajala SO (2013) Genetic analysis of maize inbred lines for tolerance to drought and low nitrogen. *JONARES* **1**, 29–36.
- Mugode L, Ha B, Kaunda A, Sikombe T, Phiri S, Mutale R, Davis C, Tanumihardjo A and De Moura FF (2014) Carotenoid retention of biofortified provitamin A maize (*Zea mays* L.) after Zambian traditional methods of milling, cooking and storage. *Journal of Agricultural and Food Chemistry* **62**, 6317–6325.
- Najafian G, Kaffashi AK and Jafar-Nezhad A (2010) Analysis of grain yield stability in hexaploid wheat genotypes grown in temperate regions of Iran using additive main effects and multiplicative interaction. *Journal of Agricultural Science and Technology* **12**, 213–222.
- Ngaboyisonga C, Njoroge K, Kirubi D and Githiri SM (2009) Effects of low nitrogen and drought on genetic parameters of grain yield and endosperm hardness of quality protein maize. *Asian Journal of Agricultural Research* **3**, 1–10.
- Owens BF, Lipka AE, Magallanes-Lundback M, Tiede T, Diepenbrock CH, Kandianis CB, Kim E, Cepela J, Mateos-Hernandez M, Buell CR, Buckler ES, DellaPenna D, Gore MA and Rocheford TR (2014) A foundation for provitamin A biofortification of maize: genome-wide association and genomic prediction models of carotenoid levels. *Genetics* **198**, 1699–1716.
- Oyekunle M and Badu-Apraku B (2014) Genetic analysis of grain yield and other traits of early – maturing maize inbreds under drought and well – watered conditions. *Journal of Agronomy and Crop Science* **200**, 92–107.
- Pillay K, Siwela M, Derera J and Veldman FJ (2014) Provitamin A carotenoids in biofortified maize and their retention during processing and preparation of South African maize foods. *Journal of Food Science and Technology* **51**, 634–644.
- Pixley K, Rojas NN, Babu R, Mutale R, Surles R and Simpungwe E (2013) Biofortification of maize with provitamin A carotenoids. In Tanumihardjo SA (ed.), *Carotenoids and Human Health*. Totowa, NJ, USA: Humana Press, pp. 271–292.
- Pswarayi A and Vivek BS (2008) Combining ability amongst CIMMYT's early maturing maize (*Zea mays* L.) germplasm under stress and non-stress conditions and identification of testers. *Euphytica* **162**, 353–362.
- SAS Institute (2012) *SAS System for Windows. Release 9.4*. Cary, NC, USA: SAS Institute Inc.
- Simpungwe E, Dhliwayo T, Palenberg M, Taleon V, Birol E, Oparinde A, Saltzman A and Diressie MT (2017) Orange maize in Zambia: crop development and delivery experience. *African Journal of Food, Agriculture, Nutrition and Development* **17**, 11973–11999. <https://doi.org/10.18697/ajfand.78.HarvestPlus08>.
- Singh RH and Chaudhary BD (1985) *Biometrical Methods in Quantitative Genetic Analysis*. New Delhi, India: Kalyani Publisher, pp. 103–157.
- Soil Survey Staff (1999) *Soil Taxonomy: A Basic System of Soil Classification for Making and Interpreting Soil Surveys*, 2nd Edn. USDA-NRCS Agriculture Handbook No. 436. Washington, DC, USA: USDA.
- Sprague GF and Tatum LA (1942) General versus specific combining ability in single crosses of corn. *Journal of the American Society of Agronomy* **34**, 923–932.
- Sultana R, Jamil S, Aslam M, Shahzad R, Fatima R, Maqbool MA and Iqbal MZ (2019) Overview of quality protein maize and molecular breeding approaches for its development. *International Journal of Biosciences* **14**, 533–542.
- Suwarno WB (2012) *Combining Ability, Association Mapping and Genomic Predictions for Provitamin A Carotenoids in Tropical Maize (Zea mays. L.)* (PhD thesis). University of Wisconsin-Madison, USA.
- Suwarno WB, Pixley KV, Palacios-Rojas N, Kaeppeler SM and Babu R (2015) Formation of heterotic groups and understanding genetic effects in a provitamin A biofortified maize breeding program. *Crop Science* **54**, 14–24.
- Teklewold A, Wegary D, Tadesse A, Tadesse B, Bantte K, Friesen D and Prasanna BM (2015) *Quality Protein Maize (QPM): A Guide to the Technology and Its Promotion in Ethiopia*. Addis Ababa, Ethiopia: CIMMYT.
- Terron A, Preciado E, Cordova H, Mickelson H and Lopez R (1997) Determinación del patron heterotico de 30 lineas de maizderivadas de la poblacion SR del CIMMYT. *Agronomy of Mesoamericana* **43**, 26–34.
- Topal A, Aydin C, Akgiin N and Babaoglu M (2004) Diallel cross analysis in durum wheat (*Triticum durum* Desf.): identification of best parents for some kernel physical features. *Field Crops Research* **87**, 1–12.
- US Institute of Medicine (2001) *Dietary Reference Intakes for Vitamin A, Vitamin K, Arsenic, Boron, Chromium, Copper, Iodine, Iron, Manganese, Molybdenum, Nickel, Silicon, Vanadium, and Zinc*. Washington, DC, USA: The National Academies Press.
- Varadaraju A and Joel J (2017) Determination of maternal effects on QPM inbred lines. *Global Journal of Bio-Science and Biotechnology* **6**, 607–611.
- Vivek BS, Krivanek AF, Palacios-Rojas N, Twumasi-Afriyie S and Diallo AO (2008) *Breeding Quality Protein Maize (QPM): Protocols for Developing QPM Cultivars*. Mexico, DF, Mexico: CIMMYT.
- Wolfe DW, Henderson DW, Hsiao TC and Alvio A (1988) Interactive water and nitrogen effects on senescence of maize. II. Photosynthetic decline and longevity of individual leaves. *Agronomy Journal* **80**, 865–870.

- Wurtzel E, Cuttriss A and Vallabhaneni R** (2012) Maize provitamin A carotenoids, current resources, and future metabolic engineering challenges. *Frontiers in Plant Science* **3**, 1–12.
- Yan W** (2001) GGE biplot – a windows application for graphical analysis of multi-environment trial data and other types of two-way data. *Agronomy Journal* **93**, 1111–1118.
- Yan W, Hunt L, Sheng Q and Szlavics Z** (2000) Cultivar evaluation and mega-environment investigation based on the GGE biplot. *Crop Science* **40**, 597–605.
- Yan W, Kang MS, Ma S, Woods S and Cornelius PL** (2007) GGE biplot vs. AMMI analysis of genotype-by-environment data. *Crop Science* **47**, 643–653.
- Yan J, Kandianis CB, Harjes CE, Bai L, Kim EH, Yang X, Skinner DJ, Fu Z, Mitchell S, Li Q, Fernandez MGS, Zaharieva M, Babu R, Fu Y, Palacios N, Li J, DellaPenna D, Brutnell T, Buckler ES, Warburton ML and Rocheford T** (2010) Rare genetic variation at *Zea mays crtRB1* increases β -carotene in maize grain. *Nature Genetics* **42**, 322–327.
- Zadoks JC, Chang TT and Konzak CF** (1974) A decimal code for growth stages of cereals. *Weed Research* **14**, 415–421.
- Zali H, Farshadfar E and Sabaghpour SH** (2011) Non-parametric analysis of phenotypic stability in chickpea (*Cicer arietinum* L.) genotypes in Iran. *Crop Breeding Journal* **1**, 85–96.